

Bioactive compounds and DPPH antioxidant activity of underutilized macroalgae (*Sargassum* spp.) from coastal water of Makassar, Indonesia

SANTIA GARDENIA WIDYASWARI^{1,2,*}, METUSALACH^{3,**}, KASMIATI³, NURSINAH AMIR³

¹Doctoral Program in Fisheries Science, Faculty of Marine Science and Fisheries, Universitas Hasanuddin. Jl. Perintis Kemerdekaan Km 10, Makassar 90245, South Sulawesi, Indonesia. Tel./fax.: +62-271-663375, *email: s.g.widyaswari@gmail.com

²Center for Quality Assurance Development of Vocational Education in the Maritime, Fisheries, Information and Communication Technology Sector. Gowa 92172, South Sulawesi, Indonesia

³Program of Fisheries Resource Utilization, Faculty of Marine Science and Fisheries, Universitas Hasanuddin. Jl. Perintis Kemerdekaan Km 10, Makassar 90245, South Sulawesi, Indonesia. Tel./fax.: +62-271-663375, **email: mminanga@hotmail.com

Manuscript received: 17 November 2023. Revision accepted: 20 January 2024.

Abstract. Widyaswari SG, Metusalach, Kasmiasi, Amir N. 2024. Bioactive compounds and DPPH antioxidant activity of underutilized macroalgae (*Sargassum* spp.) from coastal water of Makassar, Indonesia. *Biodiversitas* 25: 162-168. This study focuses on determining the chemical component and potential of two species of macroalgae (seaweed) *Sargassum* spp., i.e., *Sargassum polycystum* and *Sargassum ilicifolium* from the coastal waters of Makassar City, South Sulawesi, Indonesia. Several macroalgae are highly cultivated and exported; however, these species of *Sargassum* remain underutilized due to limited studies on their potential. Brown algae (Phaeophyceae), especially *Sargassum* spp., contain secondary metabolites known for their health-promoting properties, including antioxidant effects, inhibiting oxidative stress, lowering blood pressure, and improving the immune system. The investigation analyzes the nutritional components and bioactive substances in *Sargassum* spp. harvested from the Makassar City coastal waters, South Sulawesi, Indonesia. Nutritional content was analyzed using proximate analysis. Phytochemical screening was conducted based on the method by Harborne method. Antioxidant activity was performed using the DPPH method. Total phenolic/flavonoid content based on spectrophotometric method. The investigation reveals the primary chemical composition of *Sargassum polycystum* showing the presence of valuable secondary metabolites such as alkaloids, flavonoids, phenol hydroquinone, and reactive saponins. *Sargassum ilicifolium* contains flavonoids, phenol hydroquinone, tannins, and steroids. The IC₅₀ values for the antioxidant activity of *S. polycystum* and *S. ilicifolium* were 51.34 ppm and 51.25 ppm, respectively. The compounds in these seaweeds might be beneficial in promoting health. These findings suggest the possibility of underutilized seaweeds as a natural source of antioxidants in food and non-food industries.

Keywords: Brown algae, chemical compounds, natural antioxidant, seaweed, secondary metabolites

INTRODUCTION

The Indonesian Archipelago possesses many seaweed resources (Pambudi et al. 2010; Rimmer et al. 2021). Seaweed, also known as macroalgae, has been known for its potential benefits in the food and non-food industries (Mabeau and Fleurence 1993; Vijayabaskar and Shiyamala 2011; Tiwari and Troy 2015; Zang et al. 2020), including cosmetics (Lourenço-Lopes et al. 2020) and nutraceuticals (Shannon and Abu-Ghannam 2019). Macroalgae are an abundant source of marine biodiversity, with more than 700 species identified in Indonesia (Weber-van Bosse 1913, 1921). Some macroalgae species are cultivated and exported, such as *Kappaphycus* sp. (Valderrama et al. 2015), *Gracilaria* sp. (Nivedita and Raghunathan 2016), *Caulerpa* sp. (de Gaillande et al. 2017), *Ulva* sp. (Chemodanov et al. 2019), and *Eucheuma* sp. (Nainggolan et al. 2022). However, some other species remain underutilized due to insufficient scientific studies on their potential benefits (Moreira et al. 2022).

Certain groups of macroalgae haven't been used optimally because there haven't been sufficient scientific studies (Gazali et al. 2018; Rimmer et al. 2021). On the

other hand, extracts of brown algae have been known to contain secondary metabolites that are beneficial for health (Perumal et al. 2019; Puspita et al. 2020; Damayanti et al. 2021), including antioxidant activity that can inhibit oxidative stress (Budhiyanti et al. 2012) which can protect against degenerative diseases (Gazali et al. 2018), lowering blood pressure (Kosanić et al. 2019; Park et al. 2023), boosting the immune system (Torres et al. 2019). Some brown algae, such as *Padina*, *Turbinaria*, and *Sargassum*, have been found to have a relatively high content of beneficial compounds (Ponnan et al. 2017; Kumar et al. 2019; Jasmadi et al. 2022).

Sargassum spp. thrive in subtidal and intertidal zones, showing ecological dynamics influenced by water temperature, tidal levels, water movement, and substrate type (Afonso et al. 2019; Yip et al. 2020). These microalgae mainly attach to dead corals in shallow waters, highly exposed to ultraviolet radiation, particularly in the coastal areas of Makassar City, where they dominate the marine landscape (Mushlihah et al. 2021). The waters of the Spermonde Archipelago in South Sulawesi are impressively wealthy, hosting 199 identified macroalgae species (Verheij and Prud'homme van Reine 1993). Despite this

richness, the *Sargassum* species is still underutilized. Their life cycle was marked by detachment from substrates and subsequent transport by waves to coastal shorelines, accumulating substantial floating biomass (Sissini et al. 2017; Fidai et al. 2020; Robledo et al. 2021; Zhuang et al. 2021; Harb and Chow 2022). Attention to the potential of these species is still very limited, while interest and use of certain species of red and green macroalgae is increasing.

The underutilization of *Sargassum* could be attributed to ecological characteristics, such as exposure to ultraviolet radiation and vulnerability to wave-induced stranding. Meanwhile, red algae, especially species of *Porphyra* and *Gracilaria*, have been extensively studied and cultivated for their valuable agar and carrageenan content (Usov 1998; Bhatia et al. 2015; Liu et al. 2019; Gomes-Dias et al. 2023). Similarly, green algae, such as *Ulva* and *Enteromorpha*, have been used in biofuel production and wastewater treatment (Sode et al. 2013; Suganya et al. 2013; Bikker et al. 2016; Fouda et al. 2022). Commercial success and established cultivation practices of certain macroalgae species may overshadow the potential of underutilized varieties, such as *Sargassum*. The economic feasibility of extensively studied macroalgae may unintentionally lower the interest in exploring the potential of underutilized macroalgae.

Hence, it is crucial to determine the potential of underutilized wild macroalgae by analyzing their nutritional components and bioactive compounds. The aim of this study was to determine the nutritional components and bioactive compounds contained in *Sargassum polycystum* and *Sargassum ilicifolium* collected from the coastal waters of Makassar City, Indonesia, as well as their antioxidant activity.

MATERIALS AND METHODS

Study area and materials

Samples of *Sargassum polycystum* C.Agardh, 1824 and *Sargassum ilicifolium* (Turner) C.Agardh, 1820 were collected in April 2023 by hand-picking random sampling at the Gusung Tallang Island (5°7'19.19"S, 119°23'43.91"E), South Sulawesi, Indonesia. After collection, fresh samples were placed into plastic bags filled with ice and transported to the laboratory immediately.

Equipment used in this study includes Spectrophotometer, moisture analyzer, crucibles, a muffle furnace, a distiller, a Kjeldahl apparatus, a Soxhlet extractor, and a Buchner funnel with filter papers, centrifuge, rotary vacuum evaporator, various glassware, and micropipettes.

Proximate analysis

The sample was cleaned from debris, washed with distilled water to remove impurities and attached organisms, and dried under the sunlight (27-32°C) for approximately 48 hours. The dried sample was cut into pieces and ground into a dry powder using a chopper. Fifty g of dried *Sargassum* powder was placed into an Erlenmeyer flask

and macerated with ethanol, ethyl acetate, and hexane. The samples were macerated in each solvent for 2 × 24 hours to extract bioactive compounds in the *Sargassum* powder and then filtered using filter paper. The filtrate was concentrated with an evaporator.

The extract was analyzed for ash, water, carbohydrates, proteins, and fat contents following the AOAC Methods (Latimer 2023).

Phytochemical screening

The *Sargassum* spp. extracts underwent qualitative analysis for active compounds, including alkaloids, flavonoids, hydroquinone phenols, steroids, triterpenoids, saponins, and tannins based on the Harborne Method (Harborne 1998).

Total Phenolic Content (TPC) and Total Flavonoid Content (TFC) analysis

The quantitative analysis for Total Phenolic Content (TPC) and Total Flavonoid Content (TFC) in *Sargassum* spp. extracts were determined by the spectrophotometric method (Saeed et al. 2012). Gallic acid was used as the standard, with concentrations ranging from 0.1 to 1.0 mg/mL, then mixed with Folin-Ciocalteu reagent for TPC analysis. Following a dark incubation period of 30 minutes, sodium carbonate solution was added, and the reaction mixture was incubated for an additional 30 minutes at 25°C. Absorbance was measured at 765 nm, and the resulting data were used to construct a calibration curve and then to calculate TPC in extracts, expressed as mg Gallic Acid Equivalents (GAE) per gram extract. For TFC determination, rutin standards were similarly prepared and mixed with an aluminum chloride solution (2% to 5%), followed by a dark incubation period of 40 minutes. Absorbance was measured at 415 nm, and a calibration curve using rutin standards was used to calculate TFC in extracts, expressed as mg Rutin Equivalents (RE) per gram. The analysis was carried out in triplicate.

DPPH assay for antioxidant activity

The DPPH (2,2-diphenyl-1-picrylhydrazyl) assay for antioxidant activity follows the method by Brand-Williams et al. (1995). A fresh DPPH solution was prepared in ethanol. If necessary, the test sample or extract at concentrations ranging from 0.1 to 1.0 mg/mL was dissolved in an appropriate solvent. The DPPH solution and the sample were mixed in a 1:1 ratio and incubated in the dark for 30 to 60 minutes at room temperature. After incubation, the absorbance of the mixture was measured at 517 nm using a spectrophotometer. A higher percentage of DPPH radical scavenging activity indicated greater antioxidant activity. Ethanol as a solvent was used as a blank or negative control. For validation, a positive control containing quercetin was included in the assay. This positive control helped confirm the assay's sensitivity to detect antioxidant properties and provided a reliable reference for comparing the antioxidant activity of the test sample.

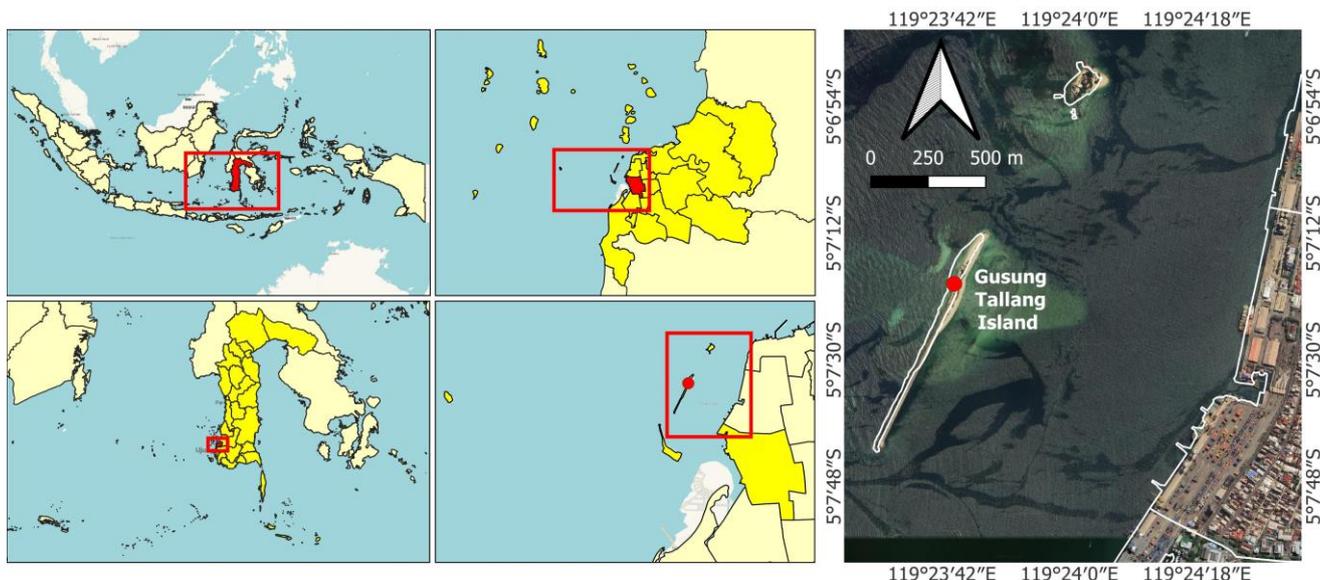


Figure 1. The Sampling site is coastal water near Makassar City, South Sulawesi, Indonesia. The red dot indicates the sampling site at Gusung Tallang Island (5° 7'19.19\"S, 119°23'43.91\"E)

Data analysis

Descriptive data analysis was done using Microsoft Excel. Results are presented as means with standard deviations. This comprehensive approach helped us thoroughly interpret and present both qualitative and quantitative results from phytochemical screening and the DPPH assay. Additionally, Pearson correlation analysis and Two-way ANOVA, using PAST 4.07b, were employed to determine correlations and differences between Total Phenolic Content (TPC), Total Flavonoid Content (TFC), and IC₅₀ values for antioxidant activity.

RESULTS AND DISCUSSION

Proximate analysis

The nutritional composition of two *Sargassum* species, i.e., *S. polycystum* and *S. ilicifolium*, is presented in Table 1. *S. ilicifolium* has a slightly higher water content (9.43%) than *S. polycystum* (8.80%), while *S. polycystum* has a higher fiber content (16.20%) than *S. ilicifolium* (12.80%). However, both species have 1.30% fat content. Water content in both species is below 10%, indicating optimal stability and a reduced risk of microbial growth. Monitoring water content is crucial for assessing self-life and determining the best storage methods to prevent microorganism growth, such as bacteria, small insects, and molds (Mannheim et al. 1994; Tun Norbrillinda et al. 2016; Obluchinskaya and Daurtseva 2020).

Sargassum polycystum has a protein content of 9.51%, while *S. ilicifolium* has 8.55%, indicating a slight difference in protein levels. Protein content in food significantly determines the quality of the food material (Friedman 1996). The crude fiber content of *S. polycystum* reached 16.20%, while *S. ilicifolium* was 12.80%. Based on the dietary guidelines of 28 g/day for adult women and 34 g/day for adult men, a daily energy intake of 2000 kcal and

2400 kcal, respectively (US Department of Agriculture (USDA) and US Department of Health and Human Services 2020), the crude fiber content of the studied macroalgae were fall within the recommended range. Dietary fiber positively affects digestion, heart disease risk, and blood sugar regulation (Nurjanah et al. 2016; Hitoie and Shimoda 2017; He et al. 2022). These results emphasize the beneficial role of crude fiber in promoting digestive health and overall well-being. The use of crude fiber should consider overall diet and individual health conditions.

Bioactive compounds

The qualitative analysis of bioactive compounds in *S. polycystum* and *S. ilicifolium* was based on color changes or precipitates formed after adding reagents (Rengasamy et al. 2020). These bioactive substances include alkaloids, flavonoids, saponins, tannins, hydroquinone phenols, and steroids/triterpenoids (Table 2). Understanding the composition of these bioactive substances is valuable for predicting their potential benefits for humans.

The *S. polycystum* contains flavonoids, hydroquinone phenol, and saponins. On the other hand, *S. ilicifolium* contains flavonoids, phenol hydroquinone, tannins, and steroids (Table 2). There are some differences in the bioactive substances in both species, even though they were collected from the same location and time. Besides species differences, the variation may be due to the age of the macroalgae and the depth of its habitat. According to previous studies, the chemical composition of macroalgae is influenced not only by species but also by factors such as maturity, reproductive type, environment, and seasons (Kumar et al. 2021; Miguel et al. 2023). In our case, the sampling was done close to the shoreline, about two meters in depth, and the macroalgae, including its roots, were collected. In particular, the leaves of *S. polycystum* (from our samples) were not dense, and the stems were soft, indicating it might be in the early growth stage.

Moreover, *S. ilicifolium* grows more densely, with broader and thicker leaves than *S. polycystum*. Light exposure and nutrients significantly influence each species' growth and chemical composition (Cronin and Hay 1996), leading to differences in bioactive substances. Even within the same species, variations in the content of bioactive compounds may occur. These variations can be due to differences in the species' age during harvest, growth conditions, and varying nutrient absorption.

Alkaloids were detected in *S. polycystum*. Alkaloids have activity as antibiotics (Chuah et al. 2016) and anti-inflammatory agents (de Souza et al. 2009), reducing pain, improving blood circulation, restoring stamina after childbirth, and preventing infections in the uterus (Zheng et al. 2017; Poole et al. 2018; Zhu et al. 2020; Sellami et al. 2022). Phenolic compounds were detected in both species. Phenolics possess anti-inflammatory properties, act as antioxidants, and contribute to carcinogen detoxification and anti-cholesterol (Michalak and Chojnacka 2015; Ravichanthiran et al. 2018; Torres et al. 2019). Saponin was detected only in *S. polycystum*. It has the potential to be an anti-inflammatory and help heal wounds. Previous studies have demonstrated that *Sargassum* spp. have antimicrobial activity (Susilowati et al. 2015; El Shafay et al. 2016; Telles et al. 2018; Abu-Khudir et al. 2021) to prevent pathogenic bacterial infection and reduce inflammation.

Antioxidant activity

Antioxidants prevent free radical autoxidation reactions in lipid oxidation (Pangestuti and Kim 2015). Antioxidant activity could be evaluated using free radical DPPH to measure radical scavenging ability by donating electrons to DPPH radicals (Table 3). The secondary metabolite analysis of both *Sargassum* species showed the presence of phenolic compounds, alkaloids, phenol hydroquinone, saponin, flavonoids, and tannins (Table 2). Phenolic compounds, distinguished by O-H bonds, showed increased antioxidant activity due to their lower dissociation bond than other groups, facilitating efficient hydrogen release and donation (Bendary et al. 2013).

The results of the antioxidant activity showed that the IC₅₀ of *S. polycystum* and *S. ilicifolium* were 51.34 and 51.25 ppm, respectively. These results suggest that both *Sargassum* species have the potential to serve as potential antioxidants (less than 100 ppm) (Table 3). Antioxidant

activity was categorized based on the IC₅₀ values obtained through the DPPH assay. A sample with an IC₅₀ value below 100 ppm is considered a potent antioxidant (Nurhasnawati et al. 2019). The IC₅₀ value reflects the concentration required to scavenge 50% DPPH radicals. The result of this study was consistent with previous research on antioxidants in *Sargassum* sp. The IC₅₀ values of *Sargassum* spp. from the coastal area of Gunung Kidul were 65.28 to 69.27 ppm (Lailatussifa et al. 2017; Sedjati et al. 2018). IC₅₀ of *Sargassum* from the coastal area of West Aceh was 68.89 ppm (Gazali et al. 2018). The results showed that *Sargassum* spp. may have the potential to be an antioxidant.

Table 1. Nutritional components of *Sargassum polycystum* and *Sargassum ilicifolium* from coastal water of Makassar City, South Sulawesi, Indonesia

Parameters	<i>S. polycystum</i>	<i>S. ilicifolium</i>
Water	8.80±0.63	9.43±0.29
Ash	28.07±0.22	38.07±0.22
Protein	9.51±0.06	8.55±0.04
Fat	1.30±0.05	1.30±0.05
Crude fiber	16.20±0.52	12.80±0.06
Carbohydrates	52.31±0.85	42.64±0.13

Table 2. Phytochemical assay of *Sargassum polycystum* and *Sargassum ilicifolium* from coastal water of Makassar City, South Sulawesi, Indonesia

Secondary metabolite	<i>S. polycystum</i>	<i>S. ilicifolium</i>	Positive test result
Alkaloid			
Mayer	-	-	White precipitate
Wagner	-	-	Brown precipitate
Dragendroff	+	-	Orange precipitate
Flavonoids	+	+	Yellowish
Phenol	+	+	Yellowish brown
hydroquinone			
Saponin	+	-	Stable foam
Tanin	-	+	White precipitate
Steroids	-	+	Yellowish brown
Triterpenoids	-	-	Yellowish brown

Table 3. Total Phenolic Content (TPC), Total Flavonoid Content (TFC), and IC₅₀ value of *Sargassum* spp. from various locations

Species	Sampling	TPC (mg GAE/g)	TFC (mg RE/g)	IC50 (ppm)	Sources
<i>Sargassum</i> sp.	Gunung Kidul	57.97	-	69.27	(Sedjati et al. 2018)
<i>Sargassum</i> sp.	West Aceh	563.22	-	68.89	(Gazali et al. 2018)
<i>Sargassum hystrix</i> J. Agardh 1847	Gunung Kidul	114.26	-	65.28	(Lailatussifa et al. 2017)
<i>Sargassum polycystum</i>	Pohuwato waters	713.00	-	77.38	(Manteu et al. 2018)
<i>Sargassum muticum</i> (Yendo) Fensholt 1955	Bangladesh	66.52	68.11	40.74	(Afrin et al. 2023)
<i>Sargassum polycystum</i>	Makassar	365.96	93.75	51.34	<i>This study</i>
<i>Sargassum ilicifolium</i>	Makassar	382.94	92.99	51.25	<i>This study</i>

The Total Phenolic Content (TPC) of *S. polycystum* and *S. ilicifolium* were 365.96 and 382.94 mg GAE/g extract, respectively. Phenolic compounds, including subclasses flavonoids and phenol hydroquinone, are known for their antioxidant capabilities. These compounds play a pivotal role in neutralizing free radicals, contributing significantly to the antioxidant potential of the sample. The increased total phenol content showed an increased antioxidant capacity, in line with the IC₅₀ values through the DPPH assay. The Total Flavonoid Content (TFC) of *S. polycystum* was 93.75 mg RE/g, and *S. ilicifolium* was 92.99 mg RE/g for *S. ilicifolium*. Based on the results of Table S1, TFC has a higher correlation with the IC₅₀ value than TPC. Table S2. showed that the TPC, TFS, and IC₅₀ values of *S. polycystum* and *S. ilicifolium* were not significantly different.

Table S1. The Pearson Correlation (Linear) matrix and plot of TPC, TCF, and IC₅₀ value.

	A (TPC)	B (TFC)	C (IC ₅₀)
A (TPC)		0.046963	0.035079
B (TFC)	0.99728		0.011883
C (IC ₅₀)	0.99848	0.99983	

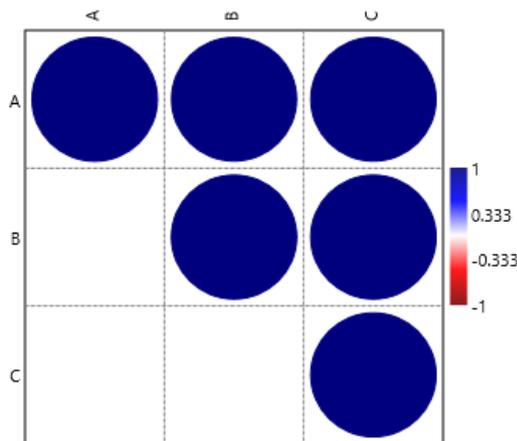


Table S2.A. Two-way ANOVA (without replication) analysis between Total Phenolic Content (TPC) and IC₅₀ value

	Sum of squares	df	Mean square	F	p (same)
Rows (between samples)	162647	5	32529.5	1.044	0.4816
Columns	274198	1	274198	8.802	0.03127
Error	155755	5	31150.9		
Total	592600	11			

Table S2.B. Two-way ANOVA (without replication) analysis between Total Flavonoid Content (TFC) and IC₅₀ value

	Sum of squares	df	Mean square	F	p (same)
Rows (between samples)	0.180625	1	0.180625	1.609	0.425
Columns	1770.31	1	1770.31	1.58E+04	0.005069
Error:	0.112225	1	0.112225		
Total:	1770.6	3			

In summary, *S. polycystum* contains alkaloids, flavonoids, phenol hydroquinone, and saponins, while *S. ilicifolium* contains flavonoids, phenol hydroquinone, tannins, and steroids. The IC₅₀ for antioxidant activity of *S. polycystum* and *S. ilicifolium* was found to be 51.34 ppm and 51.25 ppm, respectively. Further investigations using several methods to assess antioxidant activity are suggested for a comprehensive understanding of the antioxidant potential of these underutilized brown algae. The findings provide a promising starting point, and future studies should consider a broader spectrum of assays to establish a more comprehensive assessment of the antioxidant capabilities of *Sargassum* spp.

ACKNOWLEDGEMENTS

This study is part of the PhD research of the first author (SGW) at Universitas Hasanuddin, Makassar, Indonesia. The first author deeply thanks Singgih A. Putra (Universitas Hasanuddin, Indonesia) for invaluable assistance during fieldwork. The authors express sincere appreciation to two anonymous reviewers for their insightful comments and suggestions that significantly improved the quality of this work.

REFERENCES

- Abu-Khudir R, Ismail GA, Diab T. 2021. Antimicrobial, antioxidant, and anti-tumor activities of *Sargassum linearifolium* and *Cystoseira crinita* from Egyptian Mediterranean Coast. *Nutr Cancer* 73 (5): 829-844. DOI: 10.1080/01635581.2020.1764069.
- Afonso NC, Catarino MD, Silva AMS, Cardoso SM. 2019. Brown macroalgae as valuable food ingredients. *Antioxidants* 8 (9): 365. DOI: 10.3390/antiox8090365.
- Afrin F, Ahsan T, Mondal MN, Rasul MG, Afrin M, Silva AA, Yuan C, Shah AKMA. 2023. Evaluation of antioxidant and antibacterial activities of some selected seaweeds from Saint Martin's Island of Bangladesh. *Food Chem Adv* 3: 100393. DOI: 10.1016/j.focha.2023.100393.
- Bendary E, Francis RR, Ali HMG, Sarwat MI, El Hady S. 2013. Antioxidant and Structure-Activity Relationships (SARs) of some phenolic and anilines compounds. *Ann Agric Sci* 58 (2): 173-181. DOI: 10.1016/j.aos.2013.07.002.
- Bhatia S, Sharma K, Sharma A, Nagpal K, Bera T. 2015. Anti-inflammatory, analgesic and antiulcer properties of *Porphyra vietnamensis*. *Avicenna J Phytomed* 5 (1): 69-77.
- Bikker P, van Krimpen MM, van Wikselaar P, Houweling-Tan B, Scaccia N, van Hal JW, Huijgen WJJ, Cone JW, López-Contreras AM. 2016. Biorefinery of the green seaweed *Ulva lactuca* to produce animal feed, chemicals and biofuels. *J Appl Phycol* 28 (6): 3511-3525. DOI: 10.1007/s10811-016-0842-3.
- Brand-Williams W, Cuvelier ME, Berset C. 1995. Use of a free radical method to evaluate antioxidant activity. *LWT - Food Sci Technol* 28 (1): 25-30. DOI: 10.1016/S0023-6438(95)80008-5.
- Budhiyanti SA, Raharjo S, Marseno DW, Lelana IYB. 2012. Antioxidant activity of brown algae *Sargassum* species extract from the coastline of Java Island. *Am J Agric Biol Sci* 7 (3): 337-346. DOI: 10.3844/ajabssp.2012.337.346.
- Chemodanov A, Robin A, Jinjhashvily G, Yitzhak D, Liberzon A, Israel A, Golberg A. 2019. Feasibility study of *Ulva* sp. (Chlorophyta) intensive cultivation in a coastal area of the Eastern Mediterranean Sea. *Biofuels Bioprod Biorefin* 13: 864-877. DOI: 10.1002/bbb.1995.
- Chuah L-O, Shamila-Syuhada AK, Liang MT, Rosma A, Thong KL, Rusul G. 2016. Physio-chemical, microbiological properties of tempoyak and molecular characterisation of lactic acid bacteria

- isolated from tempoyak. *Food Microbiol* 58: 95-104. DOI: 10.1016/j.fm.2016.04.002.
- Cronin G, Hay ME. 1996. Effects of light and nutrient availability on the growth, secondary chemistry, and resistance to herbivory of two brown seaweeds. *Oikos* 77 (1): 93-106. DOI: 10.2307/3545589.
- Damayanti PK, Budhiyanti SA, Husni A. 2021. Antioxidant activity and consumer preference of brown algae *Sargassum hystrix* juice as a functional drink. *agriTECH* 41: 231-237. DOI: 10.22146/agritech.26665.
- de Gaillande C, Payri C, Remoissenet G, Zubia M. 2017. *Caulerpa* consumption, nutritional value and farming in the Indo-Pacific region. *J Appl Phycol* 29: 2249-2266. DOI: 10.1007/s10811-016-0912-6.
- de Souza ÉT, de Lira DP, de Queiroz AC, da Silva DJC, de Aquino AB, Mella EAC, Lorenzo VP, de Miranda GEC, de Araújo-Júnior JX, de Oliveira Chaves MC, Barbosa-Filho JM, de Athayde-Filho PF, de Oliveira Santos BV, Alexandre-Moreira MS. 2009. The antinociceptive and anti-inflammatory activities of *Caulerpin*, a bisindole alkaloid isolated from seaweeds of the Genus *Caulerpa*. *Mar Drugs* 7 (4): 689-704. DOI: 10.3390/md7040689.
- El Shafay SM, Ali SS, El-Sheekh MM. 2016. Antimicrobial activity of some seaweeds species from Red Sea, against multidrug resistant bacteria. *Egypt J Aquat Res* 42: 65-74. DOI: 10.1016/j.ejar.2015.11.006.
- Fidai YA, Dash J, Tompkins EL, Tonon T. 2020. A systematic review of floating and beach landing records of *Sargassum* beyond the Sargasso Sea. *Environ Res Commun* 2: 122001. DOI: 10.1088/2515-7620/abd109.
- Fouda A, Eid AM, Abdelkareem A, Said HA, El-Belely EF, Alkhalifah DHM, Alshallash KS, Hassan SE-D. 2022. Phyco-synthesized Zinc Oxide nanoparticles using marine macroalgae, *Ulva fasciata* Delile, characterization, antibacterial activity, photocatalysis, and tanning wastewater treatment. *Catalysts* 12: 756. DOI: 10.3390/catal12070756.
- Friedman M. 1996. Nutritional value of proteins from different food sources. A review. *J Agric Food Chem* 44: 6-29. DOI: 10.1021/jf9400167.
- Gazali M, Nurjanah N, Zamani NP. 2018. Eksplorasi senyawa bioaktif alga cokelat *Sargassum* sp. Agardh sebagai antioksidan dari pesisir barat Aceh. *Jurnal Pengolahan Hasil Perikanan Indonesia* 21 (1): 167-178. DOI: 10.17844/jphpi.v21i1.21543. [Indonesian]
- Gomes-Dias JS, Teixeira-Guedes CI, Teixeira JA, Rocha CMR. 2023. Red seaweed biorefinery: The influence of sequential extractions on the functional properties of extracted agars and porphyrans. *Intl J Biol Macromol* 257 (Pt 1): 128479. DOI: 10.1016/j.ijbiomac.2023.128479.
- Harb TB, Chow F. 2022. An overview of beach-cast seaweeds: Potential and opportunities for the valorization of underused waste biomass. *Algal Res* 62: 102643. DOI: 10.1016/j.algal.2022.102643.
- Harborne AJ. 1998. *Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis*. Springer Dordrecht, London.
- He Y, Wang B, Wen L, Wang F, Yu H, Chen D, Su X, Zhang C. 2022. Effects of dietary fiber on human health. *Food Sci Hum Wellness* 11 (1): 1-10. DOI: 10.1016/j.fshw.2021.07.001.
- Hitoe S, Shimoda H. 2017. Seaweed Fucoxanthin supplementation improves obesity parameters in mild obese Japanese subjects. *Funct Foods Health Dis* 7 (4): 246-262. DOI: 10.31989/ffhd.v7i4.333.
- Jasmadi, Kusnadi A, Kumayanjati B, Triandiza T, Pesilette RN, Ainarwowa A, Yamko AK, Pary BR, Kurnianto D. 2022. Nutritional values of *Sargassum* sp., *Ulva* sp., and *Padina* sp. from South East Molluca Island waters. *IOP Conf Ser: Earth Environ Sci* 1119: 012043. DOI: 10.1088/1755-1315/1119/1/012043.
- Kosanić M, Ranković B, Stanojković T. 2019. Brown macroalgae from the Adriatic Sea as a promising source of bioactive nutrients. *J Food Meas Charact* 13: 330-338. DOI: 10.1007/s11694-018-9948-4.
- Kumar KS, Kumari S, Singh K, Kushwaha P. 2021. Influence of Seasonal Variation on Chemical Composition and Nutritional Profiles of Macro- and Microalgae. In: Rajauria G, Yuan YV (eds). *Recent Advances in Micro and Macroalgal Processing. Food and Health Perspectives*. Wiley Blackwell, Chichester, UK. DOI: 10.1002/9781119542650.ch2.
- Kumar Y, Tarafdar A, Kumar D, Badgujar PC. 2019. Effect of Indian brown seaweed *Sargassum wightii* as a functional ingredient on the phytochemical content and antioxidant activity of coffee beverage. *J Food Sci Technol* 56: 4516-4525. DOI: 10.1007/s13197-019-03943-y.
- Lailatussifa R, Husni A, Isnansetyo A. 2017. Antioxidant activity and proximate analysis of dry powder from brown seaweed *Sargassum hystrix*. *Jurnal Perikanan Universitas Gadjah Mada* 19 (1): 29-37. DOI: 10.22146/jfs.23885.
- Latimer GW. 2023. *Official Methods of Analysis*. In: *Official Methods of Analysis of AOAC International*. Oxford University Press, New York. DOI: 10.1093/9780197610145.002.001.
- Liu Z, Gao T, Yang Y, Meng F, Zhan F, Jiang Q, Sun X. 2019. Anti-cancer activity of Porphyrin and Carrageenan from red seaweeds. *Molecules* 24 (23): 4286. DOI: 10.3390/molecules24234286.
- Lourenço-Lopes C, Fraga-Corral M, Jimenez-Lopez C, Pereira AG, Garcia-Oliveira P, Carpena M, Prieto MA, Simal-Gandara J. 2020. Metabolites from macroalgae and its applications in the cosmetic industry: A circular economy approach. *Resources* 9 (9): 101. DOI: 10.3390/resources9090101.
- Mabeau S, Fleurence J. 1993. Seaweed in food products: Biochemical and nutritional aspects. *Trends Food Sci Technol* 4 (4): 103-107. DOI: 10.1016/0924-2244(93)90091-N.
- Mannheim CH, Liu JX, Gilbert SG. 1994. Control of Water in Foods During Storage. In: Fito P, Mulet A (eds). *Water in Foods. Fundamental Aspects and Their Significance in Relation to Processing of Foods*. Pergamon, Oxford, UK. DOI: 10.1016/B978-1-85861-037-5.50036-9.
- Manteu SH, Nurjanah, Nurhayati T. 2018. Karakteristik rumput laut cokelat (*Sargassum polycystum* dan *Padina minor*) dari Perairan Pohuwato Provinsi Gorontalo. *Jurnal Pengolahan Hasil Perikanan Indonesia* 21 (3): 396-405. [Indonesian]
- Michalak I, Chojnacka K. 2015. Algae as production systems of bioactive compounds. *Eng Life Sci* 15 (2): 160-176. DOI: 10.1002/elsc.201400191.
- Miguel SP, D'Angelo C, Ribeiro MP, Simões R, Coutinho P. 2023. Chemical composition of macroalgae polysaccharides from Galician and Portugal Coasts: Seasonal variations and biological properties. *Mar Drugs* 21 (11): 589. DOI: 10.3390/md21110589.
- Moreira A, Cruz S, Marques R, Cartaxana P. 2022. The underexplored potential of green macroalgae in aquaculture. *Rev Aquac* 14 (1): 5-26. DOI: 10.1111/raq.12580.
- Mushlihah H, Amri K, Faizal A. 2021. Diversity and distribution of macroalgae to environmental conditions of Makassar City. *Jurnal Ilmu Kelautan* 7 (1): 16-26. DOI: 10.20956/jiks.v7i1.14856.
- Nainggolan PF, Arthana IW, Dewi APWK. 2022. A comparison of *Eucheuma cottonii* seaweed cultivation in monoculture and polyculture systems. *Adv Trop Biodivers Environ Sci* 6: 17-21. DOI: 10.24843/atbes.2022.v06.i01.p04.
- Nivedita S, Raghunathan C. 2016. Mariculture potential of seaweeds: With special focus on *Gracilaria* cultivation. In: Dagar JC, Sharma PC, Sharma DK, Singh AK (eds). *Innovative Saline Agriculture*. Springer India, New Delhi. DOI: 10.1007/978-81-322-2770-0_19.
- Nurhasnawati H, Sundu R, Sapri, Supriningrum R, Kuspradini H, Arung ET. 2019. Antioxidant activity, total phenolic and flavonoid content of several indigenous species of ferns in East Kalimantan, Indonesia. *Biodiversitas* 20 (2): 576-580. DOI: 10.13057/biodiv/d200238.
- Nurjanah, Nurilmala M, Hidayat T, Sudirdjo F. 2016. Characteristics of seaweed as raw materials for cosmetics. *Aquat Procedia* 7: 177-180. DOI: 10.1016/j.aqpro.2016.07.024.
- Obluchinskaya E, Daurtseva A. 2020. Effects of air drying and freezing and long-term storage on phytochemical composition of brown seaweeds. *J Appl Phycol* 32: 4235-4249. DOI: 10.1007/s10811-020-02225-x.
- Pambudi LT, Meinita MDN, Ariyati RW. 2010. Seaweed cultivation in Indonesia: Recent status. *J Mar Biosci Biotechnol* 4 (1): 6-10.
- Pangestuti R, Kim S. 2015. Seaweed proteins, peptides, and amino acids. In: Tiwari BK, Troy DJ (eds). *Seaweed Sustainability. Food and Non-Food Applications*. Academic Press, Cambridge, USA. DOI: 10.1016/B978-0-12-418697-2.00006-4.
- Park J-S, Han J-M, Shin Y-N, Park Y-S, Shin Y-R, Park S-W, Roy VC, Lee H-J, Kumagai Y, Kishimura H, Chun B-S. 2023. Exploring bioactive compounds in brown seaweeds using subcritical water: A comprehensive analysis. *Mar Drugs* 21: 328. DOI: 10.3390/md21060328.
- Perumal B, Chitra R, Maruthupandian A, Viji M. 2019. Nutritional assessment and bioactive potential of *Sargassum polycystum* C. Agardh (brown seaweed). *Indian J Geo-Mar Sci* 48 (4): 492-498.
- Poole DH, Lyons SE, Poole RK, Poore MH. 2018. Ergot alkaloids induce vasoconstriction of bovine uterine and ovarian blood vessels. *J Anim Sci* 96 (11): 4812-4822. DOI: 10.1093/jas/sky328.
- Ponnann A, Ramu K, Marudhamuthu M, Marimuthu R, Siva K, Kadarkarai M. 2017. Antibacterial, antioxidant and anticancer properties of *Turbinaria conoides* (J. Agardh) Kuetz. *Clin Phytosci* 3: 5. DOI: 10.1186/s40816-017-0042-y.
- Puspita M, Setyawidati NAR, Stiger-Pouvreau V, Vandanjon L, Widowati I, Radjasa OK, Bedoux G, Bourgougnon N. 2020. Indonesian *Sargassum* species bioprospecting: Potential applications of bioactive compounds and challenge for sustainable development. In: Bourgougnon N (eds). *Advances in Botanical Research. Seaweeds Around the World: State of Art and Perspectives*. Academic Press, Cambridge, USA. DOI: 10.1016/bs.abr.2019.12.002.

- Ravichanthiran K, Ma ZF, Zhang H, Cao Y, Wang CW, Muhammad S, Aglago EK, Zhang Y, Jin Y, Pan B. 2018. Phytochemical profile of brown rice and its nutrigenomic implications. *Antioxidants* 7 (6): 71. DOI: 10.3390/antiox7060071.
- Rengasamy KR, Mahomoodally MF, Aumeeruddy MZ, Zengin G, Xiao J, Kim DH. 2020. Bioactive compounds in seaweeds: An overview of their biological properties and safety. *Food Chem Toxicol* 135: 111013. DOI: 10.1016/j.fct.2019.111013.
- Rimmer MA, Larson S, Lapong I, Purnomo AH, Pong-Masak PR, Swanepoel L, Paul NA. 2021. Seaweed aquaculture in Indonesia contributes to social and economic aspects of livelihoods and community wellbeing. *Sustainability* 13 (19): 10946. DOI: 10.3390/su131910946.
- Robledo D, Vázquez-Delfín E, Freile-Pelegrín Y, Vázquez-Elizondo RM, Qui-Minet ZN, Salazar-Garibay A. 2021. Challenges and opportunities in relation to *Sargassum* events along the Caribbean Sea. *Front Mar Sci* 8: 699664. DOI: 10.3389/fmars.2021.699664.
- Saeed N, Khan MR, Shabbir M. 2012. Antioxidant activity, total phenolic and total flavonoid contents of whole plant extracts *Torilis leptophylla* L. *BMC Complement Altern Med* 12: 221. DOI: 10.1186/1472-6882-12-221.
- Sedjati S, Supriyanti E, Ridlo A, Soenardjo N, Santi VY. 2018. Kandungan pigmen, total fenolik dan aktivitas antioksidan *Sargassum* sp. *Jurnal Kelautan Tropis* 21 (2): 137-144. DOI: 10.14710/jkt.v21i2.3329. [Indonesian]
- Sellami M, Slimeni O, Pokrywka A, Kuvačić G, Hayes LD, Milic M, Padulo J. 2018. Herbal medicine for sports: A review. *J Int Soc Sports Nutr* 15 (14): 1-14. DOI: 10.1186/s12970-018-0218-y.
- Shannon E, Abu-Ghannam N. 2019. Seaweeds as nutraceuticals for health and nutrition. *Phycologia* 58 (5): 563-577. DOI: 10.1080/00318884.2019.1640533.
- Sissini MN, de Barros Barreto MBB, Széchy MTM et al. 2017. The floating *Sargassum* (Phaeophyceae) of the South Atlantic Ocean - likely scenarios. *Phycologia* 56 (3): 321-328. DOI: 10.2216/16-92.1.
- Sode S, Bruhn A, Balsby TJS, Larsen MM, Gotfredsen A, Rasmussen MB. 2013. Bioremediation of reject water from anaerobically digested waste water sludge with macroalgae (*Ulva lactuca*, Chlorophyta). *Bioresour Technol* 146: 426-435. DOI: 10.1016/j.biortech.2013.06.062.
- Suganya T, Gandhi NN, Renganathan S. 2013. Production of algal biodiesel from marine macroalgae *Enteromorpha compressa* by two step process: Optimization and kinetic study. *Bioresour Technol* 128: 392-400. DOI: 10.1016/j.biortech.2012.10.068.
- Susilowati R, Sabdono A, Widowati I. 2015. Isolation and characterization of bacteria associated with brown algae *Sargassum* spp. from Panjang Island and their antibacterial activities. *Procedia Environ Sci* 23: 240-246. DOI: 10.1016/j.proenv.2015.01.036.
- Telles CBS, Mendes-Aguiar C, Fidelis GP, Frasson AP, Pereira WO, Scortecci KC, Camara RBG, Nobre LTDB, Costa LS, Tasca T, Rocha HAO. 2018. Immunomodulatory effects and antimicrobial activity of heterofucans from *Sargassum filipendula*. *J Appl Phycol* 30: 569-578. DOI: 10.1007/s10811-017-1218-z.
- Tiwari BK, Troy DJ. 2015. Seaweed sustainability - food and nonfood applications. In: Tiwari BK, Troy DJ (eds). *Seaweed Sustainability. Food and Non-Food Applications*. Academic Press, Cambridge, USA. DOI: 10.1016/B978-0-12-418697-2.00001-5.
- Torres P, Santos JP, Chow F, dos Santos DYAC. 2019. A comprehensive review of traditional uses, bioactivity potential, and chemical diversity of the genus *Gracilaria* (Gracilariales, Rhodophyta). *Algal Res* 37: 288-306. DOI: 10.1016/j.algal.2018.12.009.
- Tun Norbrillinda M, Mahanom H, Nur Elyana N, Nur Intan Farina S. 2016. Optimization of spray drying process of *Sargassum muticum* color extract. *Dry Technol* 34 (14): 1735-1744. DOI: 10.1080/07373937.2016.1204550.
- US Department of Agriculture (USDA), US Department of Health and Human Services (USDHHS). 2020. *Dietary Guidelines for Americans, 2020-2025*. US Department of Agriculture (USDA), Washington, DC.
- Usov AI. 1998. Structural analysis of red seaweed galactans of agar and carrageenan groups. *Food Hydrocoll* 12 (3): 301-308. DOI: 10.1016/S0268-005X(98)00018-6.
- Valderrama D, Cai J, Hishamunda N, Ridler N, Neish IC, Hurtado AQ, Msuya FE, Krishnan M, Narayanakumar R, Kronen M, Robledo D, Gasca-Leyva E, Fraga J. 2015. The economics of *Kappaphycus* seaweed cultivation in developing countries: A comparative analysis of farming systems. *Aquac Econ Manag* 19 (2): 251-277. DOI: 10.1080/13657305.2015.1024348.
- Verheij E, Prud'homme van Reine WF. 1993. Seaweeds of the Spermonde Archipelago, SW Sulawesi, Indonesia. *Blumea - Biodivers Evol Biogeogr Plants* 37 (2): 385-510.
- Vijayabaskar P, Shiyamala V. 2011. Antibacterial activities of brown marine algae (*Sargassum wightii* and *Turbinaria ornata*) from the Gulf of Mannar Biosphere Reserve. *Adv Biol Res* 5 (2): 99-102.
- Weber-van Bosse A. 1913. *Liste des algues du Siboga I. Myxophyceae, Chlorophyceae, Phaeophyceae*. E.J. Brill, Leiden. DOI: 10.5962/bhl.title.157054.
- Weber-van Bosse A. 1921. *Liste de algues du Siboga II. Rhodophyceae*. E.J. Brill, Leiden. DOI: 10.5962/bhl.title.157054.
- Yip ZT, Quek RZB, Huang D. 2020. Historical biogeography of the widespread macroalga *Sargassum* (Fucales, Phaeophyceae). *J Phycol* 56 (2): 300-309. DOI: 10.1111/jpy.12945.
- Zang J, Xu Y, Xia W, Regenstein JM. 2020. Quality, functionality, and microbiology of fermented fish: A review. *Crit Rev Food Sci Nutr* 60 (7): 1228-1242. DOI: 10.1080/10408398.2019.1565491.
- Zheng X, Wu F, Lin X, Shen L, Feng Y. 2018. Developments in drug delivery of bioactive alkaloids derived from traditional Chinese medicine. *Drug Deliv* 25 (1): 398-416. DOI: 10.1080/10717544.2018.1431980.
- Zhu C, Liu N, Tian M, Ma L, Yang J, Lan X, Ma H, Niu J, Yu, J. 2020. Effects of alkaloids on peripheral neuropathic pain: a review. *Chin Med* 15: 106. DOI: 10.1186/s13020-020-00387-x.
- Zhuang M, Liu J, Ding X, He J, Zhao S, Wu L, Gao S, Zhao C, Liu D, Zhang J, He P. 2021. *Sargassum* blooms in the East China Sea and Yellow Sea: Formation and management. *Mar Pollut Bull* 162: 111845. DOI: 10.1016/j.marpolbul.2020.111845.