

Prope legitimate rootstocks determine the selection criteria for drought-tolerant cocoa

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Manuscript received: 2 July 2020. Revision accepted: 11 August 2020.

Abstract. *Zasari M, Wachar A, Susilo AW, Sudarsono. 2020. Prope legitimate rootstocks determine the selection criteria for drought-tolerant cocoa. Biodiversitas 21: 4067-4075.* Drought tolerant cocoa seedlings are needed for the success of cocoa cultivation under drought conditions. Prope legitimate rootstock is feasible to be used in the selection of drought-tolerant cocoa seedlings. This objective of this study was to determine the selection criteria and tolerance of prope legitimate cocoa rootstocks in response to drought stress. This study was conducted in the greenhouse of Kaliwining experimental garden, Indonesian Center for Coffee and Cocoa Research, Jember, Indonesia. Plant materials were arranged in a split-plot design with 2 factors and 6 replications. The main plot was the soil moisture content, i.e 100%, and 25%, while the subplots were 13 prope legitimate cocoa rootstocks. The result showed that root fresh weight was the best characters for the selection of drought tolerance in prope legitimate cocoa rootstock. Based on the value of stress susceptible index and clustering analysis divided the tested prope legitimate rootstock into two groups. The group for tolerant genotype was consisted of ICCRI 03, ICS 60, TSH 858, KKM 22, KW 641, KW 516, and MCC 02, while the sensitive genotype group was consisted of Sul 01, Sul 03, Sca 06, KEE 02, KW 617, and Sul 02.

Keywords: Cocoa, prope legitimate rootstock, selection criteria, drought tolerance

INTRODUCTION

Climate instability is a serious challenge to the productivity and stability of agricultural crops (Waseem et al. 2011; Kyei-Mensah et al. 2019). Extreme rainfall fluctuations allow the plant to experience a water deficit (Mishraa and Cherkauer 2010; Bari et al. 2016; Medina and Laliberte 2017) which impacts on its final production (Zlatev and Lidon 2012). Cocoa is a very sensitive plant to drought stress (Carr and Lockwood 2011; Amos and Thompson 2015; De Almeida 2016; Gateau-Rey et al. 2018; Santhyami et al. 2018; Yoroba et al. 2019). Drought stress causes seedling mortality, fruit and seed size reduction, and the increase of pest and disease attacks on cocoa (Atayese et al. 2013; Longe and Oyekale 2013; Santos et al. 2014; Ofori et al. 2015; Gateau-Rey et al. 2018).

Drought condition is a threat to cocoa cultivation. Drought tolerant cacao seedling is needed to overcome the threat of drought (Cazares, et al. 2010) as well as the expansion of cocoa plantations to the marginal area with drought conditions as the limiting factor (Setyawan and Susilo 2017; Setyawan et al. 2018). Exploration has been carried out through a number of studies related to the characterization and selection to obtain drought-tolerant cacao seedling (Atayese et al. 2013; Dos Santos et al. 2016; De Almeida et al. 2016; Zakariyya et al. 2017; Setyawan et al. 2018; Zakaryya and Indradewa 2018a, 2018b).

Drought tolerant cacao seedling can be produced through generative and vegetative propagation methods. As

one of popular vegetative methods, grafting has been widely used by cocoa farmers in Malaysia, Philippines, or even Indonesia (Sodré and Gomes 2019). In addition to its easiness to practice, the grafting technique produced superior seedling because it can accommodate the combination of two good characteristics from two different parents, i.e rootstock and scion (Pranowo and Wardiana 2016). Rootstock is the lower part of seedling, while scion was the upper part that combined through grafting techniques. Rootstock was combined with the scion in the upper part. The rootstock should be prepared from any varieties with a good adaptation to the lack of water, nutrition, and other environmental stress (Lopez-Ortega et al. 2016; Warschefsky et al. 2016). In general, the propagation technique of cacao rootstock uses a controlled crossbreed seed. However, the use of open pollinated seeds is seemed to be more feasible for rootstock production (Susilo 2015; Zakariyya et al. 2017).

Prope legitimate seed is a term for any seed that produced by mother plant who receive random pollen from one or several clones that grow in the same block. This seed can be used as the source of genotype diversity that is required during the selection of drought-tolerant rootstock. The study of tolerance capability of prope legitimate rootstock should be conducted in the dry field instead of in vitro condition (Atayese et al. 2012; Dos Santos et al. 2016; De Almeida et al. 2016; Setyawan et al. 2018). The success to obtain prope legitimate rootstock is highly dependent on the accuracy of selection criteria, however, there is still

limited studies regarding this issue. Therefore, this study aimed to determine the selection criteria and tolerance of prope legitimate cocoa seedling under drought stress conditions.

MATERIALS AND METHODS

Experimental site, time and plant materials

This study was conducted at the greenhouse of Kaliwining experimental garden, Indonesian Center for Coffee and Cocoa Research, Jember, East Java, Indonesia. The study was conducted from July to December 2018. The average temperature and relative humidity of the greenhouse during the study period were 25-30°C and 59.6-89.3%, respectively. Plant materials were prepared from the propagation of prope legitimate seed clones namely KW 516, KW 617, KW 641, ICCRI 03, TSH 858, Sca 06, MCC 02, KEE 02, KKM 22, ICS 60, Sul 01, Sul 02, and Sul 03. All tested plants were grown in polybag with a size of 15 cm x 25 cm containing 1300 g soil media until 2 months old.

Experimental method

The study used a split-plot design with 2 factors and 6 replications. The first factor as the main plot was the soil moisture content, i.e. 100% and 25%, while the second factor as the subplot was 13 prope legitimate cocoa rootstocks, i.e. KW 516, KW 617, KW 641, ICCRI 03, TSH 858, Sca 06, MCC 02, KEE 02, KKM 22, ICS 60, Sul 01, Sul 02, and Sul 03. The moisture content was controlled manually by adding a certain amount of water to the planting medium every 5 days until the seedlings were 5 months old (Setyawan et al. 2018). Determination of moisture content following the Gravimetric method (Dobriyal et al. 2012). Several variables that observed at 3 months after treatment were the percent of survived seedlings; the plant height and the root length that was measured using the bar; the stem diameter that was measured using calipers; the number of leaves that were calculated manually; the weight of plant canopy and root (both in term of dry and wet basis) that was weighed by using a digital balance; the area of leaf and root were measured by using ImageJ software, and the root volume was measured following the volumetric method in beaker glass.

Other observed variables were stomatal density and relative water content. Stomatal density was determined through the imprint technique. The sample was selected leaf on the 3rd position from the apical bud. The abaxial part of the leaf was smeared with transparent nail polish as wide as ± 0.5 cm and the allow it to dry. The dry nail polish is assumed to be attached to the epidermis and stomata cells. Next, the dry nail polish was gently removed using sticky tape and then placed on the object-glass for further observation under the microscope. Stomata were observed with a lens magnification for about 10 times and then calculated using software called Image Raster. The relative water content was calculated from the following equation (Pizard et al. 2011) as follow:

$$\text{RWC (\%)} = [(\text{Wf} - \text{Wd}) / (\text{Ws} - \text{Wd})] \times 100$$

Where: RWC is relative water content (%), Wf is the leaves fresh weight, Wd is the leaves dry weight, Ws is the leaves turgid weight.

Proline content also measured in present experiment. Proline was measured by using the method of Abraham et al. (2010). The sample was prepared in form of 0.5 g leaves. The leaf was crushed with mortar, then add by 10 ml 3% sulfosalicylic acid and filtered with filter paper. The mixture of filtrate with 2 ml ninhydrin acid and 2 ml glacial acetic acid in a test tube was heated at 96°C for 60 minutes and then terminated by cooling the solution inside the ice water for about 5 minutes. The product was extracted with 4 ml toluene (99.5%) and then shaken with vortex for 15-20 seconds to form two layers (chromoform). The red top layer containing proline was taken with a pipette and then the absorbance was measured using a spectrophotometer at a wavelength of 520 nm. The content of proline was determined by reading the standard of pure proline.

Data analysis

Data were processed through analysis of variance by using R software version 3.44. The homogeneity test was performed by using the F-test at a 5% confidence level. Determination of selection criteria referred to variables that showed significant effect on the variety of interactions between genotypes and the environment. Data from the selected variables were processed by relative decrease analysis to distinguish genotype responses. Analysis of the component was carried out to assess the estimation value of inheritance and variability (Syukur et al. 2011). Genotype tolerance was analyzed using stress sensitivity index that was calculated by the formula: $\text{SSI} = (1 - Y_s / Y_p) / (1 - \bar{Y}_s / \bar{Y}_p)$ (Fisher and Maurer 1978), Y_s = average specific genotype of drought stress conditions, Y_p = average of an optimum condition genotype, \bar{Y}_s = average of all drought stress genotypes, and \bar{Y}_p = average of all optimum condition genotypes. The grouping of rootstock genotypes was performed by using a heatmap clustergram analysis.

RESULTS AND DISCUSSION

Agronomical characters as the main selection criteria

Selection was an important step in efforts to improve plant genetic quality. Selection based on the mean value of population was the most suitable method for family-level selection in the annual crops (Mayo 1980; Padi et al. 2013). Prope legitimate rootstock was obtained from the seed multiplication with the same female parent. Tolerant characters of cocoa rootstock could be predicted through identification of specific characters as selection characters (Santos et al. 2014; Ofori et al. 2015; Fang and Xiong 2015). Selection character was determined by using certain approach that refers to the influence of various interactions between genotype and environment and relative decrease. Characters with significant interaction variance showed the different intergenotype responses to stress (Mohammadi and Abdulahi 2017; Anshori et al. 2018a; Anshori et al.

2019). The analysis of variance in Table 1 showed various interaction variance in all observed characters. The characters of stem fresh weight, stem dry weight, root fresh weight, dry weight, and canopy fresh weight showed a significant interaction variance. Thus, it was the indicator that five mentioned characters were likely to be desired selection criteria.

The tolerance level of plants to drought stress could be estimated based on the magnitude of the relative decrease in various plant growth characters. The calculation of relative decrease was performed only for significant characters. The results of relative decrease analysis (Table 2) showed that there was a decrease in the average value of stem fresh weight, stem dry weight, root fresh weight, root dry weight, and canopy fresh weight in all observed genotypes in response to drought stress. Relative decrease indicated the response level of plant genotype to stress (De Leon et al. 2015; Anshori et al. 2018b, 2019).

Determination of the right selection criteria could improve the efficiency and the accuracy of the selection process (Santos et al. 2014; Ofori et al. 2015; Fang and Xiong 2015; Dos Santos 2016). The right selection criteria were indicated by the possibility to be inherited to the offspring. The proper selection criteria should have a heritability value. The estimation of heritability aimed to evaluate the suitability between genetic potential and phenotypic performance of selected plants. In general, there were two types of heritability values, namely broad sense heritability (h_{bs}^2) and narrow-sense heritability (h_{ns}^2) (Mayo 1980; Islami et al. 2015; Baloch et al. 2016). The broad-sense heritability represented the magnitude of the role of genetic to the phenotypic aspect, so it was often used to determine the selection character (Rubiyo and Sudarsono 2011; Islam et al. 2015; Dos Santos et al. 2016).

The result of the variance component (Table 3) showed that there was the root fresh weight character with the highest estimation on the broad inheritance value (h_{bs}^2). The broad inheritance value on root fresh weight character in present study was categorized as high level (Syukur et al. 2011), so that the character of root fresh weight was effectively used in determining selection criteria. The

higher broad sense inheritance value might lead to the higher effect of genetics resulting in the phenotypic performances; in this case, the root fresh weight (Mayo 1980; Rubiyo and Sudarsono 2011; Dos Santos et al. 2016). The phenotypic expression of this root character was influenced either by genetic and environmental factors (Padi et al. 2013; Geonaga et al. 2015). Relatively controlled environmental conditions on both climatic and soil factors allowed the genetic factor to influence the maximum expression of root fresh weight.

Genetic variability described the genetic background of the tested genotypes, while the variability of phenotype is oriented towards the physical appearance of the genotypes tested. The level of genetic variability was determined by the genetic variance (σ_g^2) and the deviation standard of genetic variance (σ_{σ_g}), as well as for the variability of phenotype. Both of the genetic variance and phenotype variance of stem fresh weight, root fresh weight, and root dry weight were smaller than twice of standard deviation of genetic and phenotype variances, respectively (Table 3). Thus, the variability of root fresh weight characters was relatively narrow. As predicted, the narrow genetic diversity was directly followed by the narrow phenotypic diversity, because the phenotypic diversity was affected not only by environment but also genetic (Syukur et al. 2010).

In general, the selection process paid attention to characters with a broad genetic variability, however, the determination of selection character should be adjusted to the desired objectives (Susilo 2015; Devy et al. 2018). The improvement of rooting properties would affect the tolerance of the cocoa rootstock. Thus, root fresh weight was appropriate characters chosen as the selection criteria for cocoa prope legitimate rootstock against drought stress. The root weight character was an observed variable that indicated the root growth. One of several approaches to assess plant adaptation was the evaluation of the plant ability to maximize the rooting system under lack of water condition (Atayase et al. 2013; Placide et al. 2014; Setyawan et al. 2018).

Table 1. Analysis of variance of agronomical characters of prope legitimate cocoa rootstocks under drought stress condition

Source	PSS (%)	SD (cm)	PH (cm)	NL (leaf)	LA (cm)	FWL (g)	DWL (g)	FWS (g)
Water content	0.00**	0.00**	0.00**	0.00**	0.00**	0.00**	0.00**	0.00**
Genotype	0.21 ^{tn}	0.00**	0.00**	0.00**	0.00**	0.39 ^{tn}	0.59 ^{tn}	0.00**
Water content.Genotype	0.29 ^{tn}	0.23 ^{tn}	0.97 ^{tn}	0.46 ^{tn}	0.07 ^{tn}	0.13 ^{tn}	0.35 ^{tn}	0.00**
CV (%)	9.08	5.57	9.75	8.54	8.89	18.85	19.39	13.95
	DWS (g)	FWR (g)	DWR (g)	RA (cm)	RL (cm)	RV (cm)	FWC (g)	DWC (g)
Water content	0.00**	0.00**	0.00**	0.03*	0.50 ^{tn}	0.00**	0.00**	0.00**
Genotype	0.05 ^{tn}	0.00**	0.00**	0.00**	0.00**	0.00**	0.00**	0.00**
Water content.Genotype	0.01*	0.03*	0.02*	0.05 ^{tn}	0.52 ^{tn}	0.12 ^{tn}	0.00**	0.12 ^{tn}
CV (%)	19.09	15.82	23.35	16.61	8.35	23.57	13.03	16.01

Note: *significantly different at α 5%, ** significantly different at α 1%, ^{tn} not significantly different, EN: experimental number, CV: coefficient of variance, PSS: the percentage of survived seedling, SD: stem diameter, PH: plant height, NL: the number of leaves, LA: the leaf area, FWL: the fresh weight of leaf, DWL: the dry weight of leaf, FWS: the fresh weight of stem, DWS: the dry weight of stem, FWR: the fresh weight of root, DWR: the dry weight of root, RA: the root area, RL: the root length, RV: the root volume, FWC: the fresh weight of plant canopy, DWC: the dry weight of plant canopy

Table 2. The relative decrease of stem fresh weight stem dry weight, root fresh weight root dry weight, and canopy fresh weight of prope legitimate cacao rootstock genotypes under drought stress condition

Genotype	Stem fresh weight (g)				Stem dry weight (g)				Root fresh weight (g)				Root dry weight (g)				Canopy fresh weight (g)			
	Environment			RD	Environment			RD	Environment			RD	Environment			RD	Environment			RD
	Opt	Sts	Delta	(%)	Opt	Stress	Delta	(%)	Opt	Stress	Delta	(%)	Opt	Stress	Delta	(%)	Opt	Stress	Delta	(%)
KW 516	10.82	5.38	5.44	50.32	4.90	2.39	2.51	51.30	4.19	2.76	1.43	34.21	1.38	1.13	0.25	18.04	20.88	9.30	11.57	55.44
KW 617	9.37	5.35	4.02	42.92	4.15	2.24	1.91	46.02	4.01	2.15	1.85	46.26	1.36	0.77	0.59	43.68	18.13	10.66	7.46	41.18
KW 641	8.41	5.47	2.94	35.02	3.85	2.37	1.49	38.59	3.99	2.83	1.16	29.07	1.26	1.24	0.02	1.23	16.77	9.08	7.70	45.89
ICCRI 03	8.17	5.24	2.94	35.90	3.94	1.98	1.96	49.70	3.00	2.30	0.70	23.20	1.10	0.86	0.24	21.49	16.83	9.49	7.34	43.60
TSH 858	8.81	5.55	3.26	36.96	3.78	2.43	1.34	35.57	3.52	2.53	0.99	28.24	1.09	0.82	0.27	24.58	17.05	10.25	6.80	39.91
Sca 06	7.99	4.85	3.13	39.25	4.01	1.85	2.16	53.87	3.66	2.42	1.24	33.96	1.20	0.80	0.40	33.39	16.60	8.39	8.21	49.45
KEE 02	9.34	5.48	3.86	41.35	4.77	2.19	2.59	54.19	3.99	2.47	1.52	38.13	1.46	0.95	0.50	34.42	18.68	8.85	9.83	52.61
KKM 22	7.76	5.93	1.83	23.63	3.59	2.43	1.16	32.26	3.77	2.90	0.86	22.91	1.27	1.14	0.13	10.43	15.81	9.64	6.17	39.01
MCC 02	9.93	6.26	3.67	36.99	4.56	2.46	2.10	46.09	4.39	3.15	1.24	28.27	1.64	1.12	0.52	31.87	18.79	10.30	8.48	45.15
ICS 60	7.19	5.14	2.06	28.56	3.82	2.14	1.68	44.02	3.26	2.39	0.88	26.84	1.17	0.90	0.27	23.18	15.21	9.61	5.60	36.83
Sul 01	9.51	4.70	4.81	50.54	4.78	2.14	2.64	55.30	3.92	2.52	1.41	35.85	1.47	0.82	0.64	43.85	18.72	8.86	9.86	52.67
Sul 02	9.01	4.61	4.40	48.87	4.41	1.81	2.59	58.89	4.07	2.08	1.99	48.96	1.29	0.76	0.54	41.52	17.59	8.28	9.31	52.92
Sul 03	8.62	4.96	3.66	42.51	4.45	1.76	2.69	60.48	4.25	2.41	1.84	43.26	1.62	0.82	0.80	49.29	17.34	9.66	7.67	44.26

Note: opt: the optimum condition, sts: the stressed condition, delta: the difference of mean obtained from the results of optimum condition minus the stressed condition, RD: the relative decrease of observed variables

Physiological and biochemical characters of prope legitimate cocoa rootstocks

Drought stress-induced various plant responses in terms of morphological, physiological, and biochemical characters (Borem et al. 2012). The physiological and biochemical responses of plants at seedling stage could be used to predict the tolerance to drought stress (Santos et al. 2014; Ofori et al. 2015; Fang and Xiong 2015). Relative water content (Lugojan and Ciulca 2011; Anjum et al. 2011a; Ghobadi et al. 2011; Zakaryya et al. 2017), stomatal density (Zakaryya et al. 2017; Lahive et al. 2018), and proline content (Anjum et al. 2011b; Kishor and Sreenivasulu 2014; Zakaryya and Indradewa 2018a) frequently reported to be selection index for determining plant tolerance to drought stress.

The result showed that relative water content, stomatal density, and leaf proline content had a significant interaction variance in response to drought stress (Table 4). Thus, these characters indicated the various responses under drought stress conditions.

Based on the analysis of relative decreases in Table 5, there was various reduction in term of relative water content and stomatal density of rootstock in response to drought stress, even some genotype did not show a decrease in relative water content and stomata density. The decrease of relative water content and stomata density was indicated the plant responses to water stress (Ghobadi et al. 2011; Zakaryya et al. 2017). All prope legitimate rootstock genotypes showed an elevated leaf proline content (Table 5). In response to osmotic stress such salinity and drought, most plants accumulated proline as natural osmolyte and also for preventing the cell from reactive oxygen species (ROS) (Anjum et al. 2011b; Polavarapu et al. 2014; Zakariyya and Indradewa 2018a).

Estimation of index and prope legitimate cocoa rootstock tolerance

Evaluation of drought tolerance characteristics of genotypes can be predicted using susceptible stress index (SSI). SSI values are used to measure the yield stability due to changes in the environment related to the mechanism of resistance and genotype sensitivity (Khayatnezhad and Gholamin 2012; Anshori et al. 2018b). The results of analysis based on the SSI value of root fresh weight and root dry weight characters (Table 6), showed that the prope legitimate cocoa rootstock genotypes were divided into two groups, namely medium tolerant and susceptible to drought stress. Genotypes that had a SSI value of $0.5 < SSI \leq 1$ were indicated to be medium tolerant, while groups of genotypes that had a SSI value of > 1.00 were sensitive (Fisher and Maurer 1978).

Grouping was used to determine the similarity among tested plant genotypes. There was a need to know which characters that classified the plant population into the group. This information could be used to ease the selection process. There was a number of grouping method, and one of them was clustergram analysis. Clustergram analysis was a multivariate analysis by combining several clusters into a flat dimension. The contrast color could ease to understand in which characters specified to a certain group (Lee et al. 2016). This analysis had carried out on morphological characters (Zimisuhara et al. 2015; Anshori et al. 2018a). As the consequence, the simple and easy understanding related to grouping of genotypes could increase the selection effectiveness (Yuan et al. 2016).

Table 3. The component of variance, heritability, and deviation standard of genetic characters in prope legitimate cacao rootstock genotypes under drought stress condition

Characters	Variance and Standard deviation				Heritability h_{bs}^2	Variability	
	σ_g^2	σ_{σ_g}	σ_p^2	σ_{σ_p}		$2(\sigma_{\sigma_g})$	$2(\sigma_{\sigma_p})$
Stem fresh weight	0.07	0.25	0.33	1.52	0.21	0.49	3.04
Root fresh weight	0.04	0.06	0.08	1.49	0.50	0.13	2.99
Root dry weight	0.01	0.01	0.02	1.11	0.27	0.03	2.23

Note: σ_g^2 : variance of genetic, σ_p^2 : variance of phenotype, h_{bs}^2 : the broad-sense heritability, σ_{σ_g} = deviation standard of genetic variance, σ_{σ_p} = deviation standard of phenotype variance

Table 4. Analysis of variance of physiological and biochemical characters of prope legitimate cocoa rootstocks under drought stress condition

Source	Relative water content (%)	Stomatal density (stomata/cm ²)	Leaf proline content (µmol/g)
Water content	0.00**	0.84 ^{tn}	0.00**
Genotype	0.73 ^{tn}	0.00**	0.00**
Water content.Genotype	0.00**	0.00**	0.00**
CV (%)	7.36	11.64	10.27 ^{tr}

Note: *significantly different at α 5%, ** significantly different at α 1%, ^{tn} not significantly different, ^{tr}: transformation data (log+1.5), CV: coefficient of variance

Table 5. The effect of drought stress on the relative water content, stomatal density and leaf proline content in prope legitimate cocoa rootstocks

Genotype	Relative water content (%)				Stomatal density (stomata/cm ²)				Leaf proline content (μmol/g)			
	Environment			RD	Environment			RD	Environment			RI
	Opt	Sts	delta	(%)	Opt	Sts	delta	(%)	Opt	Sts	delta	(%)
KW 516	65.50	54.16	11.35	17.32	121.85	112.66	9.19	7.54	5.88	35.11	35.86	83.60
KW 617	55.86	65.50	-9.65	0.00	107.46	121.85	-14.40	0.00	4.37	213.16	217.07	97.99
KW641	68.65	55.86	12.79	18.63	99.97	107.46	-7.48	0.00	4.09	126.96	88.85	95.40
ICCRI 03	54.48	68.65	-14.17	0.00	103.37	99.97	3.40	3.29	2.52	99.41	136.45	98.15
TSH 858	66.78	54.48	12.29	18.41	113.50	103.37	10.13	8.92	1.50	45.04	38.60	96.11
Sca 06	57.17	66.78	-9.61	0.00	119.76	113.50	6.26	5.23	3.48	28.51	34.08	89.79
KEE 02	63.60	57.17	6.43	10.12	127.78	119.76	8.02	6.27	4.15	122.89	124.02	96.65
KKM 22	47.46	63.60	-16.14	0.00	121.48	127.78	-6.29	0.00	5.74	98.85	99.86	94.25
MCC 02	69.38	47.46	21.92	31.60	119.49	121.48	-1.99	0.00	2.43	145.31	145.86	98.33
ICS 60	53.35	69.38	-16.03	0.00	123.81	119.49	4.31	3.48	2.10	103.17	55.18	96.19
Sul 01	67.43	53.35	14.08	20.89	92.34	123.81	-31.47	0.00	3.17	18.30	18.64	83.00
Sul 02	49.29	67.43	-18.14	0.00	108.68	92.34	16.35	15.04	1.76	142.41	142.45	98.76
Sul 03	68.29	49.29	19.00	27.82	109.17	108.68	0.49	0.45	11.69	44.00	46.55	74.87

Note: opt: the optimum condition, sts: the stressed condition, delta: the difference of mean obtained from the results of optimum condition minus the stressed condition, RD: the relative decrease of observed variables, RI: the relative increase of observed variables

Table 6. Tolerance characteristic of prope legitimate cocoa rootstock on the drought sensitivity index on root fresh weight and root dry weight characters

Genotype	Root fresh weight (g)				Root dry weight (g)			
	Optimum	Stress	SSI	Criteria	Optimum	Stress	SSI	Criteria
KW 516	4.19	2.76	1.00	Medium tolerant	1.38	1.13	0.60	Medium tolerant
KW 617	4.01	2.15	1.35	Susceptible	1.36	0.77	1.46	Susceptible
KW 641	3.99	2.83	0.85	Medium tolerant	1.26	1.25	0.04	Tolerant
ICCRI 03	3.00	2.30	0.68	Medium tolerant	1.10	0.86	0.72	Medium tolerant
TSH 858	3.52	2.53	0.83	Medium tolerant	1.09	0.82	0.82	Medium tolerant
Sca 06	3.66	2.42	0.99	Medium tolerant	1.20	0.80	1.12	Susceptible
KEE 02	3.99	2.47	1.11	Susceptible	1.45	0.95	1.15	Susceptible
KKM 22	3.77	2.90	0.67	Medium tolerant	1.27	1.13	0.35	Tolerant
MCC 02	4.39	3.15	0.83	Medium tolerant	1.64	1.12	1.07	Medium tolerant
ICS 60	3.26	2.39	0.78	Medium tolerant	1.17	0.90	0.78	Medium tolerant
Sul 01	3.92	2.52	1.05	Susceptible	1.46	0.82	1.47	Susceptible
Sul 02	4.07	2.08	1.43	Susceptible	1.29	0.76	1.39	Susceptible
Sul 03	4.25	2.41	1.26	Susceptible	1.62	0.82	1.65	Susceptible

Note: SSI: susceptible stress index

The results of the clustergram analysis (Figure 1) showed that there was two groups of prope legitimate cocoa rootstocks. This grouping was made based on the pattern of color intensity on the selection characters supported by the susceptible stress index. The first group was divided into subgroup 1 which consists of several genotypes, namely ICCRI 03, ICS 60, and TSH 858, while subgroup 2 which consists of KKM 22, KW 641, KW 516, and MCC 02. The second group was divided into subgroup 1 which consists of two genotypes, namely Sul 01 and Sul 3, while subgroup 2 which consists of Sca 06, KEE 02, KW 617, and Sul 02. The first group was the group that was indicated to be tolerant rootstock genotypes. The second group was the group that was indicated to be sensitive

rootstock genotypes. In general, the first group showed the relative decrease trend of root fresh weight and root dry weight was lower than the second group. Genotypes responded to water deficit conditions by decreasing root weight (Paez-garcia et al. 2015). The lower relative decrease, the more tolerant the stress. Tolerant genotype was characterized by the plant ability to survive and remain productive under stress conditions (Cazares et al. 2010; Rukundu et al. 2014). Tolerant genotypes usually possessed a deep rooting system with greater number of root hair in drought conditions. The large root volumes could be able to absorb more water so that they could survive in the water shortages condition (Paez-garcia et al. 2015; Setyawan et al. 2018).

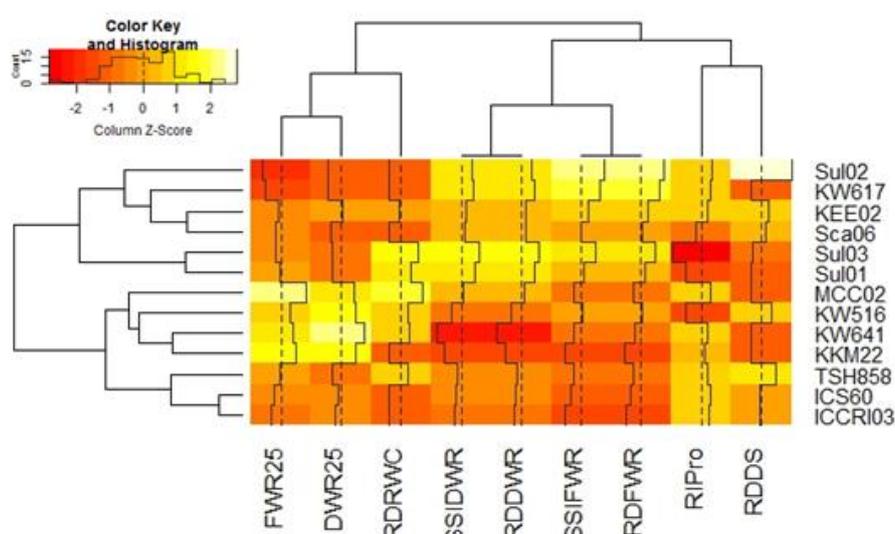


Figure 1. The clustergram heatmap of 13 prope legitimate cocoa rootstock genotypes under drought condition based on 9 observed variables, i.e FWR25: root fresh weight in stressed condition, DWR25: root dry weight in stressed condition, RDRWC: relative decrease of water content, SSIDWR: Susceptible stress index of root dry weight, RDDWR: relative decrease of root dry weight, SSIFWR: Susceptible stress index of root fresh weight, RDFWR: relative decrease of root fresh weight, RIPro: relative increase of proline content, and RDDS: relative decrease of stomatal density

The reduction of relative water content and stomatal density including increased in proline content of tolerant and sensitive genotypes were various responses (Figure 1). In general, Genotypes responded to water deficit conditions by decreasing relative water content (Medeiros et al. 2012; Zakaryya et al. 2017) and stomatal density (Zhao et al. 2015; Zakaryya et al. 2017). The plant water status does not only depend on the water availability in plant tissue, but also the stomatal conductivity for opening and closing activities (Ghobadi et al. 2011; Anjum et al. 2011b). The reduction of relative water content and stomatal density were highly influenced by genetic factors (Yang and Mio 2010). Increased proline production was plant adaptation strategy to survive in drought conditions (Medeiros et al. 2012). Proline accumulation was the plant response to drought stress in order to reduce cell damage (Anjum et al. 2011b; Polavarapu et al. 2014). The level of proline accumulation depended on genotype, growing stage, stress level, and other environmental conditions (Wang et al. 2015).

In conclusion, the root fresh weight was the best criterion to select the drought-tolerant prope legitimate cocoa rootstock. The drought-tolerant prope legitimate cocoa rootstocks were ICCRI 03, ICS 60, TSH 858, KKM 22, KW 641, KW 516, and MCC 02, while the sensitive prope legitimate cocoa rootstocks were Sul 01, Sul 03, Sca 06, KEE 02, KW 617, and Sul 02.

ACKNOWLEDGEMENTS

This work was supported by the Ministry of Research, Technology, and Higher Education of Indonesia through

the scheme of postgraduate scholarship program. We also thanked the Indonesian Center for Coffee and Cocoa Research for providing the plant materials.

REFERENCES

- Anshori MF, Purwoko BS, Dewi IS, Ardie SW, Suwarno WB, Safitri H. 2018a. Determination of selection criteria for screening of rice genotypes for salinity tolerance. *SABRAO J Breed Genet* 50 (3): 279-294.
- Anshori MF, Purwoko BS, Dewi IS, Ardie SW, Suwarno WB, Safitri H. 2018b. Heritability, characterization, and clustergram analysis of doubled haploid rice lines derived from anther culture. *Jurnal Agronomi Indonesia* 46 (2): 119-125.
- Anshori MF, Purwoko BS, Dewi IS, Ardie SW, Suwarno WB, Safitri H. 2019. Selection index based on multivariate analysis for selecting double-haploid rice lines in lowland saline prone area. *SABRAO J Breed Genet* 51 (2): 161-174.
- Amos TT, Thompson TO. 2015. Climate change and the cocoa production in the tropical rain forest ecological zone of Ondo State, Nigeria. *J Environ Earth Sci* 5 (1): 36-41.
- Anjum SA, Wang XXL, Saleem MF, Man C, Lei W. 2011a. Morphological, physiological and biochemical responses of plants to drought stress. *Review. Afr J Agric Res* 6 (9): 2026-2032.
- Anjum SA, Xie Xy, Wang Lc, Saleem MF, Man C, Lei W. 2011b. Review: Morphological, physiological and biochemical responses of plants to drought stress. *Afr J Agric Res* 6 (9): 026-2032.
- Atayese M, Olaiya AO, Adedeji AR, Hammed LA. 2013. Evaluation of the three Cocoa varieties for Drought tolerance in Nigeria. *Nigerian J Horticult Sci* 17: 117-187.
- Bari A, Damania AB, Mackay M, Dayanandan S. 2016. Applied Mathematics and Omics to Assess Crop Genetic Resources for Climate Change Adaptive Traits. CRC Press, Boca Raton, FL.
- Baloch AW, Bhatti SM, Baloch M, Jogi QD, Kandhro MN. 2016. Correlation analysis of various metric traits with grain yield and heritability estimation in rice genotypes. *Pak J Agric Eng Vet Sci* 32: 136-142.
- Borem A, Ramalho MAP, Fritsche-Neto R. 2012. Abiotic stresses: Challenges for plant breeding in the coming decades: *Plant Breeding*

- for Abiotic Stress Tolerance. Fritsche-Neto R, Borem A (eds.). Springer-Verlag, Berlin.
- Carr MKV, Lockwood G. 2011. The water relations and irrigation requirements of cocoa (*Theobroma cacao* L.). *Expl Agric* 47 (4): 653-676.
- Cazares BX, Elenes FF, Medrano RR. 2010. Drought tolerance in crop plants. *Am J Plant Physiol* 5 (5): 241-256.
- De Almeida J, Tezara W, Herrera A. 2016. Physiological responses to drought and experimental water deficit and waterlogging of four clones of cacao (*Theobroma cacao* L.) selected for cultivation in Venezuela. *Agric Water Manag* 171: 80-88.
- De Leon, T.B, S. Linscombe, G. Gregorio, P.K. Subudhi. 2015. Genetic variation in southern USA rice genotypes for seeding salinity tolerance. *Front Plant Sci* 8 (374): 1-13.
- Devy L, Anita-sari I, Susilo AW, Wachjar A, Sobir. 2018. Genetic diversity and indirect selection of fine cacao (*Theobroma cacao*) based on bean color. *Biodiversitas* 19 (6): 2385-2392.
- Dobriyal P, Qureshi A, Badola R, Hussain SA. 2012. A review of the methods available for estimating soil moisture and its implications for water resource management. *J Hydrol* 458-459 (2012): 110-117.
- Dos Santos EA, de Almeida AAF, Ahnert D, da Silva Branco MC, Valle RR, Baligar VC. 2016. Diallel analysis and growth parameters as selection tools for drought tolerance in young *Theobroma cacao* plants. *PLoS ONE* 11 (8): e0160647. DOI: 10.1371/journal.pone.0160647.
- Fang Y, Xiong L. 2015. General mechanisms of drought response and their application in drought resistance improvement in plants. *Review. Mol Life Sci* 72: 673-689.
- Fischer, R.A, Maurer R.. 1978. Drought resistance in spring wheat cultivars. I. Grain yield response. *Aust J Agric Res* 29: 897-907.
- Galmes J, Flexas J, Save R, Medrano H. 2007. Water relations and stomatal characteristics of Mediterranean plants with different growth forms and leaf habits: Responses to water stress and recovery. *Plant Soil* 290: 139-155.
- Gateau-Rey L, Tanner EVJ, Rapidel B, Marelli J-P, Royaert S. 2018. Climate change could threaten cocoa production: Effects of 2015-16 El Niño-related drought on cocoa agroforests in Bahia, Brazil. *PLoS ONE* 13 (7): e0200454. DOI: 10.1371/journal.pone.0200454.
- Ghobadi M, Khosravi S, Kahrizi D, Shirvani F. 2011. Study of water relations, chlorophyll and their correlations with grain yield in wheat (*Triticum aestivum* L.) genotypes. *Intl J Biol Biomol Agric Food Biotechnol Eng* 5 (6): 353-356.
- Goenaga R, Gultinan M, Maximova S, Seguíne E, Irizarry H. 2015. Yield performance and bean quality traits of cacao propagated by grafting and somatic embryo-derived cuttings. *HortScience* 50 (3): 358-362.
- Islam MA, Raffi SA, Hossain MA, Hasan AK. 2015. Analysis of genetic variability, heritability, and genetic advance for yield and yield associated traits in some promising advanced lines of rice. *Progr Agric* 26: 26-31.
- Kishor PBK, Sreenivasulu N. 2014. Is proline accumulation per se correlated with stress tolerance or is proline homeostasis a more critical issue?. *Mini Review. Plant Cell Environ* 37: 300-311.
- Khayatnezhad, M, Gholamin R. 2012. The effect of drought stress on leaf chlorophyll content and stress resistance in maize cultivars (Zea mays). *Afr J Microbiol Res* 6 (12): 2844-2848.
- Kyei-Mensah C, Kyerematen R, Adu-Acheampong S. 2019. Impact of rainfall variability on crop production within the Worobong Ecological Area of Fantekwa District, Ghana. *Adv Agric* 2019: 7930127. DOI: 10.1155/2019/7930127.
- Lahive F, Hadley P, Daymond AJ. 2018. The impact of elevated CO₂ and water deficit stress on growth and photosynthesis of juvenile cacao (*Theobroma cacao* L.). *Photosynthetica* 56 (3): 911-920.
- Lee, JE, Recker M, Bowers AJ, Yuan M. 2016. Hierarchical cluster analysis heatmaps and pattern analysis: an approach for visualizing learning management system interaction data. *Proceeding of The 9th International Conference on Educational Data Mining. North Carolina, USA, 29 June-2 July 2016.*
- Longe OA, Oyekale AS. 2013. Assessment of climate change vulnerability and adaptation among smallholder cocoa farmers in Osun State, Nigeria. *Life Sci J* 10 (2): 757-763.
- Longenberger PS, Smith CW, Thaxton PS, McMichael BL. 2006. Development of a screening method for drought tolerance in cotton seedlings. *Crop Sci* 46: 2104-2110.
- Lopez-Ortega G, García-Montiel F, Bayo-Canha A, Frutos-Ruiz C, Frutos-Tomás D. 2016. Rootstock effects on the growth, yield and fruit quality of sweet cherry cv. "Newstar" in the growing conditions of the Region of Murcia. *Scientia Horticulturae* 198: 326-335.
- Lugojan C, Ciulca S. 2011. Analysis of excised leaves water loss in winter wheat. *J Horticult For Biotechnol* 15 (2): 178-182.
- Mayo O. 1980. *The Theory of Plant Breeding*. Oxford University Press. New York.
- Medeiros DB, Ciriaco E, Rafael H, Santos B, Pacheco CM, Musser S, Jurema R, Custódio M. 2012. Physiological and biochemical responses to drought stress in Barbados Cherry. *Braz Soc Plant Physiol* 24: 181-192.
- Medina V, Laliberté B. 2017. A review of research on the effects of drought and temperature stress and increased CO₂ on *Theobroma cacao* L. and the role of genetic diversity to address climate change. *Bioversity International, Costa Rica.*
- Mishra V, Cherkauer KA. 2010. Retrospective droughts in the crop growing season: Implications to corn and soybean yield in the Midwestern United States. *Agric For Meteorol* 150: 1030-1045.
- Mohammadi R, Abdulahi A. 2017. Evaluation of durum wheat genotypes based on drought tolerance indices under different levels of drought stress. *J Agric Sci* 62 (1): 1-14
- Ofori A, Padi FK, Acheanpong K, Lowor S. 2015. Genetic variation and relationship of traits related to drought tolerance in cocoa (*Theobroma cacao* L.) under shade and no-shade conditions in Ghana. *Euphytica*. 201: 411-421.
- Ostenson I, Püttsepp Ü, Biel C, Alberton O, Bakker MR, Löhmus K, Metcalfe D, Olsthoorn AFM, Pronk A, Vanguelova E, Weih M, Brunner I. 2007. All aspects of plant biology: specific root length as an indicator of environmental change. *Plant Biosyst* 141: 426-442.
- Padi FK, Adu-Gyamfi P, Akpertey A, Arthur A, Ofori A. 2013. Differential response of cocoa (*Theobroma cacao*) families to field establishment stress. *Plant Breed* 132 (2): 229-236.
- Paez-garcía A, Motes CM, Chen R, Blancafor EB, Monteros MJ, Samuel T, Noble R, Parkway SN. 2015. Root traits and phenotyping strategies for plant improvement. *Plants* 4: 334-355.
- Pirzad A, Shakiba MR, Zehtab-Salmasi S, Mohammadi SA, Darvishzadeh R, Samadi A. 2011. Effect of water stress on leaf relative water content, chlorophyll, proline and soluble carbohydrates in *Matricaria chamomilla* L. *Med Plants Res* 5 (12): 2483-2488.
- Placide R, Hirut GB, Stephan N, Fekadu B. 2014. Assessment of drought stress tolerance in root and tuber crops. *Afr J Plant Sci* 8 (4): 214-224.
- Polavarapu B, Kishor K, Sreenivasulu N. 2014. Review: Is proline accumulation per se correlated with stress tolerance or is proline homeostasis a more critical issue?. *Plant Cell Environ* 37: 300-311.
- Pranowo D, Wardiana E. 2016. Kompatibilitas lima klon unggul kakao sebagai batang atas dengan batang bawah progeny *Half-Sib* klon Sulawesi 01. *Jurnal Tanaman Industri dan Penyegar* 3: 29-36. [Indonesian]
- Rubiyo, Sudarsono. 2011. Pendugaan Parameter Genetik Ketahanan Tanaman Kakao terhadap penyakit busuk buah. *Buletin RISTRI* 2 (3): 391-404. [Indonesian]
- Rukundu P, Betaw HG, Ngailo S, Balcha F. 2014. Assessment of drought stress tolerance in root and tuber crops. *Review. Afr J Plants Sci* 8 (4): 214-224.
- Santos IC, de Almeida AAF, Ahnert D, da Conceic AS, Pirovani CP, Pires JL, Valle RR, Baligar VC. 2014. Physiological and biochemical responses of *Theobroma cacao* L. genotypes to soil water deficit. *PLoS ONE* 9(12): e115746. DOI: 10.1371/journal.pone.0115746
- Santhyami, Basukriadi A, Patria MP, Abdulhadi R. 2018. The comparison of aboveground C-stock between cacao-based agroforestry system and cacao monoculture practice in West Sumatra, Indonesia. *Biodiversitas* 19 (2): 472-479.
- Setyawan B, Puspitasari N, Susilo AW, Anita-Sari I. 2018. Rootstock characteristics of three combinations of *Theobroma cacao* L. crosses on different water availability. *Pelita Perkebunan* 34 (3): 137-145.
- Setyawan B, Susilo AW. 2017. Selection of prospective drought-tolerant cocoa hybrids based on additive main effect and multiplicative interaction analyses. *Pelita Perkebunan* 33 (2): 89-96.
- Sodre GA, Gomes ARS. 2019. Cocoa propagation, technologies for production of seedlings. *Revista Brasilia Frutic Jaboticabal* 41: 1-22.
- Susilo AW. 2015. *Bahan Tanam Kakao: Kakao Sejarah, Botani, Proses Produksi, Pengolahan, Perdagangan*. Pusat Penelitian Kopi dan Kakao Indonesia. Gadjah Mada University Press, Yogyakarta. [Indonesian]
- Syukur M, Sujiprihati S, Yunianti R, Nida K. 2010. Pendugaan komponen genetik, heritabilitas dan korelasi untuk menentukan kriteria seleksi cabai (*Capsicum annum* L.) populasi 15. *J Hort Indonesia* 1 (2): 74-80. [Indonesian]

- Syukur M, Sujiprihati S, Yuniarti R, Kusumah DA. 2011. Pendugaan ragam genetik dan heritabilitas karakter komponen hasil beberapa genotipe cabai. *Agrivigor* 10 (2): 148-156. [Indonesian]
- Wang H, Tang X, Wang H, Shao H-B. 2015. Proline accumulation and metabolism-related genes expression profiles in *Kosteletzkya virginica* seedlings under salt stress. *Front Plant Sci* 792: 1-9. DOI: 10.3389/fpls.2015.00792.
- Warschefsky EJ, Klein LL, Frank MH, Chitwood DH, Londo JP, von Wettberg EJB, Miller AJ. 2016. Rootstocks: Diversity, domestication, and impacts on shoot phenotypes. *Trends Plant Sci* 21 (5): 418-437.
- Waseem M, Ali A, Tahir M, Nadeem A, Ayub M, Tanveer A, Ahmad R, Hussain M. 2011. Mechanism of drought tolerance in plant and its management through different methods. *Agric Sci* 5 (1): 10-25.
- Xu Z, Zhou G. 2008. Responses of leaf stomatal density to water status and its relationship with photosynthesis in a grass. *J Exp Bot* 59 (12): 3317-3325.
- Yang F, Miao LF (2010). Adaptive responses to progressive drought stress in two poplar species originating from different altitudes. *Silva Fennica* 44: 23-37.
- Yuan J, Murphy A, Koeyer DD, Lague M, Bizimungu B. 2016. Effectiveness of the field selection parameters on potato yield in Atlantic Canada. *Can J Plant Sci* 96: 701-710.
- Yoroba F, Kouassi BK, Diawara A, Yapo LAM, Kouadio K, Tiemoko DT, Kouadio YK, Kone ID, Assamoi P. 2019. Evaluation of rainfall and temperature conditions for a perennial crop in tropical wetland: A case study of cocoa in Côte d'Ivoire. *Adv Meteorol* 2019: 9405939. DOI: 10.1155/2019/9405939
- Zakariyya F, Setiyawan B, Susilo AW. 2017. Stomatal, proline, and leaf water status characters of some cocoa clones (*Theobroma cacao* L.) on prolonged dry season. *Pelita Perkebunan* 33 (1): 109-117.
- Zakariyya F, Indradewa D. 2018a. Biochemical changes of three cocoa clones (*Theobroma cacao* L.) under drought stress. *Ilmu Pertanian (Agricultural Science)* 3 (2): 82-88.
- Zakariyya F, Indradewa D. 2018b. Drought stress affecting growth and some physiological characters of three cocoa clones at seedling phase. *Pelita Perkebunan* 34 (3): 156-165.
- Zhao W, Sun Y, Kjelgren R, Liu X. 2015. Response of stomatal density and bound gas exchange in leaves of maize to soil water deficit. *Acta Physiol Plant* 37: 1704. DOI: 10.1007/s11738-014-1704-8.
- Zlatev Z, Lidon FC. 2012. An overview on drought-induced changes in plant growth, water relations and photosynthesis. *J Food Agric* 24 (1): 57-72.
- Zimisuhara B, Valdiani A, Shaharuddin NA, Qamaruzzaman F, Maziah M. 2015. Structure and principal component analyses reveal an intervarietal fusion in Malaysia mistletoe fig (*Ficus deltoidea* Jack) population. *Intl J Mol Sci* 16: 14369-14394.