

Sequence variation of bovine mitochondrial ND-5 between haplotypes of composite and Hereford Breeds of beef cattle

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ABSTRACT

The aims of the study were to: Investigate polymorphisms in the ND-5 region of bovine mitochondrial DNA in the composite and purebred Hereford herds from the Wokalup selection experiment, sequencing and compare the sequences between haplotypes and published sequence from Genbank. A total of 194 Hereford and 235 composite breed cattle from Wokalup Research Station were used in this study. The mitochondrial DNA was extracted using Wizard genomic DNA purification system from Promega. ND-5 fragment of mitochondrial DNA was amplified using PCR and continued with RFLP. Each haplotypes were sequenced. PCR products of each haplotype were cloned into pCR II, transformed, colonies selection, plasmid DNA extraction continued with cycle sequencing. Polymorphisms were found in both breeds of cattle in ND-5 region of mitochondrial DNA by PCR-RFLP analysis. Sequencing analysis confirmed the RFLPs data.

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Key words: ND-5 mitochondrial DNA, PCR-RFLP, sequencing, polymorphism.

INTRODUCTION

Genetic diversity is the basis for livestock breeding (Buis, Oldenbroek and Vanderwerf, 1994), because it is used as a starting point for the improvement of breeds by artificial selection. Understanding the extent and pattern of genetic variability among breeds may help in the development of more rational breeding programs and is a prerequisite to the informed conservation of genetic resources (Kidd *et al.*, 1974). It thus the information on genetic diversity and genetic relationships among cattle breeds may be very useful in cattle breeding programs.

Advanced techniques of molecular biology have provided the opportunity to study genetic diversity within and among breeds at the single gene level (Baker and Manwell, 1991; Loftus *et al.*, 1994b; MacHugh *et al.*, 1994; Moazamigoudarzi *et al.*, 1997). Many DNA markers, either of genomic DNA or cytoplasmic DNA, have been generated

recently by utilizing molecular techniques. Mitochondrial DNA, a genetic information located outside nuclear cell, has become an area of interest for studying the maternal inheritance of many traits in livestock, as well as in population genetic studies. Avise (1994) has extensively reviewed the advantages of using mitochondrial DNA, a cytoplasmically inherited DNA, as a genetic marker in population studies. Mitochondrial DNA polymorphism has been reported within and between breeds of cattle (Bhat, Mishra and Bhat, 1990; Ron *et al.*, 1990; Sutarno and Lymbery, 1997; Tanaka *et al.*, 1995) and recently, Loftus *et al.* (1994a) used mtDNA polymorphisms to study the phylogeny of different breeds of cattle from Europe, Asia and Africa. For the application of studying mitochondrial DNA, except in phylogenetic studies, variation of mitochondrial DNA has been suggested to significantly correlate with fertility trait in beef cattle (Sutarno *et al.*, 2002).

MATERIALS AND METHODS

Blood collection

Blood was collected by venepuncture into a 50ml tube containing 2.5ml of 200mM EDTA as anticoagulant. 10ml of this blood was aspirated and stored at -70°C for future reference. White blood cells were then isolated from the remaining 40ml of blood.

Isolation of white blood cells

Whole blood was dispensed into centrifuge tubes, and then spun at about 1500g for 15-20 minutes. The buffy coat was removed with a pipette, transferred to 10ml or 20 ml centrifuge tubes, topped up with TE-1 buffer (10mM Tris, 1mM EDTA, pH 8) and centrifuged at 2000g for 10-15 minutes. The pellet was resuspended in 1ml of TE-2 buffer (10mM Tris, 1mM EDTA and 100mM NaCl, pH 8.0), transferred to 1ml Nunc storage tube, and frozen at -84°C.

Extraction of mitochondrial DNA from white blood cells

Mitochondrial DNA was extracted using the Wizard Minipreps DNA Purification System (Promega, Madison, USA). Mitochondrial pellets were prepared according to published methods (Welter, Dooley and Blin, 1989).

PCR-RFLP

All PCR amplification reactions were performed in an Omnigene thermocycler machine. The reactions were performed in a 50 ml reaction mix consisting of 200 ng of template DNA, 0.15 μ M each of the oligonucleotide primers, 200 μ M each dNTPs, 2 mM MgCl₂, 10x buffer and 1.5 units Taq DNA polymerase (Biotech, Australia) in 0.6 μ l PCR reaction tube.

PCR amplification of mitochondrial ND-5

The ND-5 region, a part of the gene coding for NADH dehydrogenase sub-unit 5 (ND-5) were amplified by PCR, using primers ND-L / ND-R are described below:

ND-5 Primers:

ND-L: 5'-ATCCGTTGGTCTTAGGAACC-3'

ND-R: 5'-TTGCGGTTACAAGGATGAGC-3'

All amplification reactions were performed in a 50 l reaction mix consisting of 200 ng of template DNA, 0.15 μ M each of the oligonucleotide primers, 200 μ M each dNTPs,

2 mM MgCl₂, 10x buffer and 1.5 units Taq DNA polymerase (Biotech, Australia). Negative controls (lacking template DNA) were included in all reactions, and produced no products.

PCR amplification conditions for the mitochondrial ND-5 were as follows: one preliminary denaturation reaction was set at 94 °C for 6 minutes, followed by 30 amplification cycles, in which each cycle consisted of strand denaturation at 94 °C for 45 sec, annealing at 58 °C for 45 sec and primer extension at 72 °C for 1 min, followed by a final polymerisation at 72 °C for 6 minutes.

Extraction of plasmid DNA

DNA containing inserts were extracted from selected liquid culture of overnight incubation using Wizard Plus Minipreps DNA Purification system (Promega, Madison, USA).

Producing PCR products for cloning

The conditions for PCR amplification of the ND-5 regions were the same as with PCR for RFLP analysis, except the final step. To make sure all amplified DNA is double stranded with 3' A-overhangs, the final extension step was set for 10 minutes.

Cloning into pCR II

Fresh PCR products were electrophoresed on a 1 % low melting agarose gel. Bands were excised and purified using Wizard PCR Preps (Promega). The purified fragments were then precipitated using 99 % ethanol and 3 M Na acetate. DNA content was measured using a fluorometer.

The ratio between vector and insert is very important in determining the efficiency of ligation. Based on the fluorometer reading and the length (base pair) of PCR products, the amount of PCR product needed for ligation can be estimated using the formula given in the Invitrogen instruction manual. 1.1 ng of ND-5 (A and B haplotype) were separately put in the microcentrifuge containing 1 μ l of 10 x ligation buffer, 2 μ l of pCR II vector (25 ng/ μ l), 4.9 μ l sterile water and 1 μ l T4 DNA ligase (4.0 Weiss units). The ligation reaction was incubated at 14°C overnight.

Transformation

Tubes containing the ligation reactions were centrifuged briefly and placed on ice.

Vials of 0.5 M β -mercaptoethanol (β -ME) and a 50 μ l vial of frozen One Shot competent cells (one for each ligation/transformation) were thawed on ice. 2 μ l of 0.5 M β -ME was pipetted into each vial of the competent cells and mixed by stirring gently with the pipette tip. 2 μ l of each ligation reaction was then pipetted into the competent cells and mixed by stirring with the pipette tip. The tubes were then incubated on ice for 30 minutes followed by heat shock for exactly 30 seconds in the 42°C water bath, and then placed on ice for 2 minutes. 450 μ l of SOC medium (2 % tryptone, 0.5 % yeast extract, 10 mM NaCl, 2.5 mM KCl, 10 mM MgSO₄, 20 mM glucose, from: Invitrogen, San Diego, CA) were added into each tube. The tubes were then placed in a rotary shaking at 37°C for 1 hour at 225 rpm. The tubes with the transformed cells were placed on ice. 50 ml and 200 ml from each transformation tube was spread on separate, labeled Luria Bertani (LB) agar (1 % tryptone, 0.5 % yeast extract, 1 % NaCl; pH 7.0) plates containing 50 μ g/ml kanamycin and X-Gal. The plates were then left at room temperature for 1 hour for the liquid to be absorbed by the agar, inverted and placed in a 37°C room for 18 hours.

Blue white selection of colonies

After 18 hours incubation, blue and white colonies appeared on each agar plate. The white colonies contained inserts and were selected for further culture. Selection was done from the plates containing 50 μ l spreaded transformation mix. By using a sterile toothpick, a white colony was put into a 30 ml sterile plastic tube containing 5 ml liquid culture (LB broth). Ten colonies for each plate were taken into 10 culture tubes, and incubated in a rotary shaker (225 rpm) at 37°C overnight.

PCR selection

After 4 hours incubation, 20 μ l of each culture tube were taken for PCR selection, and the tubes were returned immediately into the rotary shaker. Each 20 ml sample was put into a separate microcentrifuge tube containing 1% Triton X in TE buffer pH 8.0. The tubes were then boiled at 97°C for 10 minutes followed by centrifugation at 14,000 rpm for 5 minutes. 5 μ l of the supernatant from each tube was transferred into 20 μ l PCR reaction using M13

forward and reverse primers. The PCR products were then electrophoresed on a 1 % agarose gel to check the incorporation of the insert in the amplified fragment.

Plasmid DNA extraction from liquid culture

DNA containing ND-5 (A and B) inserts were then extracted from selected liquid cultures using DNA Minipreps Plus (Promega). The amount of extracted DNA was then measured using a fluorometer to correctly estimate the amount of DNA used for cycle sequencing. The extension products were precipitated using 99 % ethanol and 3 M sodium acetate before sequencing.

Purification of extension products

Extension products were purified using the QIAquick PCR Purification Kit (Qiagen, Germany). The product were then stored at -20°C prior to sequencing.

Cycle sequencing

Mitochondrial ND-5 were sequenced using a dye-labeled terminator cycle sequencing kit supplied by Applied Biosystems. Aliquots of 2 μ l of double stranded DNA extracted from the cloned vector containing an insert of the D-loop were added into a microcentrifuge tube consisting of 8.0 μ l terminator ready reaction mix, 3.2 pmole primer and dH₂O adjusted to final volume of 20 μ l followed by cycle sequencing reaction.

The cycle sequencing reaction was performed in a thermal cycler as follows: (1) Twenty five cycles of rapid thermal ramp to 96°C, 96°C for 30 seconds, rapid thermal ramp to 50°C, 50°C for 15 seconds, rapid thermal ramp to 60°C, 60°C for 4 minutes; (2) Rapid thermal ramp to 4°C and hold.

RESULTS

Restriction site polymorphisms in mitochondrial ND-5

Polymorphism resulted from amplified ND-5 fragment digested using SpeI enzyme, one of six enzymes used in this study, was presented in Figure 1.

The PCR products of the ND-5 were digested using 6 enzymes listed in Table 1, and the cleavage patterns of those enzymes are shown in Table 1.



Figure 1. Gel photograph of an ethidium bromide stained agarose gel showing mitochondrial ND-5 polymorphism detected by PCR-RFLP using *SpeI*.

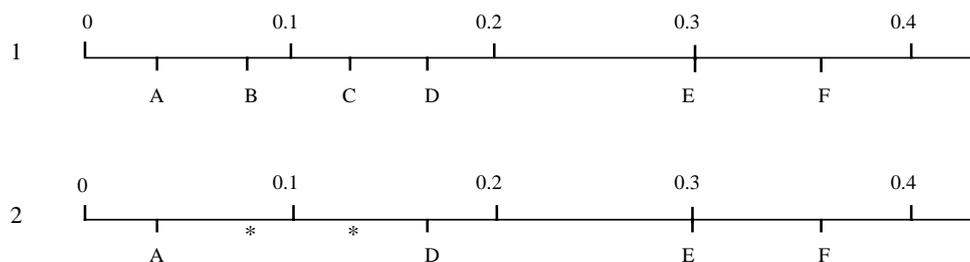


Figure 2. Restriction maps of ND-5. The enzymes are: *BstXI* (A), *SpeI* (B), *Hind III* (C), *SnaBI*(D) *DraI* (E) and *EcoRI* (F). The restriction sites are according to the sequence of (Anderson *et al.*, 1982). Polymorphic sites are indicated by *. The two haplotypes coded by the alleles in Table 3.6 are: 1. (AAAAAA); and 2. (ABAABA).

Table 1. Restriction sites of 6 enzymes on 453-bp ND-5 fragment of mitochondrial DNA amplified by PCR.

Enzyme	Allele	No of Restriction site	Fragment size (kb)
<i>BstXI</i>	A	1	0.42, 0.04
<i>SpeI</i>	A	1	0.38, 0.08
	B	0	0.46
<i>HindIII</i>	A	1	0.33, 0.13
	B	0	0.46
<i>SnaBI</i>	A	1	0.29, 0.17
<i>DraI</i>	A	1	0.30, 0.16
<i>EcoRI</i>	A	1	0.36, 0.10

Two polymorphic sites were detected on ND-5 fragment using *SpeI* and *HindIII*. Two haplotypes resulted, and the map of restriction sites of those haplotype were presented in Figure 2.

Sequencing results

Alignment of mitochondrial ND-5 sequences of different genotypes with the gene sequence from genebank (Anderson *et al.*, 1982). Sequence similarities or differences are shown with or without asterik (*).

DISCUSSION

Recent developments in molecular techniques have resulted in an abundance of data on genetic polymorphisms from DNA analysis. These data will provide us with a better understanding of the nature of genetic variation within and between cattle breeds. PCR-RFLP analysis can detect the same type of polymorphisms as traditional RFLP analysis, but without the need for Southern

CLUSTAL W (1.4) multiple sequence alignment.

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ND5-GENE BANK      ATCCGTTGGTCTTAGGAACCAAAAAATTGGTGCAACTCCAAATAAAAAGTAATAAACATAT
SUT-ND-5-A         ATCCGTTGGTCTTAGGAACCAAAAAATTGGTGCAACTCCAAATAAAAAGTAATAAACATAT
SUT-ND-5-B         ATCCGTTGGTCTTAGGAACCAAAAAATTGGTGCAACTCCAAATAAAAAGTAATAAACATAT
*****

ND5-GENE BANK      TCTCCTCACTCTCACTAGTTACTTTACTCTTACTACTATACCCATTATAATAATAAGCT
SUT-ND-5-A         TCTCCTCACTCTCACTAGTTACTTTACTCTTACTACTATACCCATTATAATAATAAGCT
SUT-ND-5-B         TCTCCTCACTCTCACTGGTTACTTTACTCTTACTACTATACCCATTATAATAATAAGCC
*****

ND5-GENE BANK      TTAACACCTACAAACCTTCCAACCTACCCACTCTACGTAAAAACAGCTATCTCATACGCCT
SUT-ND-5-A         TTAACACCTACAAACCTTCCAACCTACCCACTCTACGTAAAAACAGCTATCTCATACGCCT
SUT-ND-5-B         TTAACACCTACAAACCTTCCAACCTACCCACTCTACGTAAAAACAGCTATCTCATATGCCT
*****

ND5-GENE BANK      TCATTACCAGCATAATTCCCACAATAATATTTATCCACTCAGGCCAAGAACTAATTATTT
SUT-ND-5-A         TCATTACCAGCATAATTCCCACAATAATATTTATCCACTCAGGCCAAGAACTAATTATTT
SUT-ND-5-B         TCATTACCAGCATAATTCCCACAATAATATTTATCCACTCAGGCCAAGAACTAATTATTT
*****

ND5-GENE BANK      CAAACTGACACTGACTAACCATCCAAACTCTTAAATTTATCCCTCAGCTTTAAAATAGACT
SUT-ND-5-A         CAAACTGACACTGACTAACCATCCAAACTCTTAAATTTATCCCTCAGCTTTAAAATAGACT
SUT-ND-5-B         CAAACTGACACTGACTAACCATCCAAACTCTTAAATTTATCCCTCAGCTTTAAAATAGACT
*****

ND5-GENE BANK      ATTTCTCAATAATATTTATCCCAGTAGCACTATTCGTCACATGATCTATTATAGAATTCT
SUT-ND-5-A         ATTTCTCAATAATATTTATCCCAGTAGCACTATTCGTCACATGATCTATTATAGAATTCT
SUT-ND-5-B         ATTTCTCAATAATATTTATCCCAGTAGCACTATTCGTCACATGATCTATTATAGAATTCT
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blotting (Cushwa and Medrano, 1996), thus decreasing the time taken and increasing sensitivity (Weber and May, 1989). This method is therefore very useful for the study of genetic variation. Moreover, automatic sequence analysis, a relatively new method of analysis, has enable researchers to detect any mutation in the DNA the level of gene.

Polymorphisms were found in the mitochondrial ND-5 region with *SpeI* and *HindIII*, confirming previous reports (Suzuki, Kemp and Teale, 1993; Watanabe *et al.*, 1985b; Watanabe *et al.*, 1989). New polymorphisms have been reported in the ND-5 by PCR-RFLP analysis (Sutarno and Lymbery, 1997), and this polymorphism was reported to be significantly affect fertility trait in beef cattle (Sutarno *et al.*, 2002). Compared to the standard bovine sequence of mitochondrial DNA (Anderson *et al.*, 1982), by aligning the sequencing results with previous sequence from Genbank, these variant sequences have lost an *SpeI* and *HindIII* sites in mitochondrial ND-5.

Genetic variation within breeds is important and its study has become a subject of interest in livestock species, as it has many applications in animal breeding and genetics. Archibald (1983) listed applications such as the identification of animals and parentage testing, gene mapping and identifying markers for performance traits. Since all phenotypic characters are influenced by the genetic information carried by DNA, DNA variation may be correlated with variation in performance traits. This idea is the basis for marker assisted selection (MAS), which has aroused much interest in recent years (Schwerin *et al.*, 1995; Soller, 1994). Genetic variation, measured at the DNA level, can also be used as a check on the level of genetic variation in quantitative traits maintained within breeds. This will be extremely valuable in maintaining pools of genetic diversity, which artificial selection often acts to reduce (Archibald, 1983).

Another important application of genetic variation between breeds is to predict the

crosses between breeds that will produce crossbreed offspring with maximum heterosis. Much more attention has been paid in recent years to the utilization of heterosis in beef cattle and other livestock species. However, because there are so many breeds that could be used for crossbreeding, it is impossible to experimentally cross and compare all breeds. The ability to predict the magnitude of heterosis effects between breeds is therefore crucially needed (Goddard and Ahmed, 1982). Since previous reports have indicated that heterosis does not increase in any simple way with increasing performance differences between parental populations (Robertson and Reeve, 1955), and relationships between heterosis and marker gene distance have been found (Ehiobu, Goddard and Taylor, 1990), the study of genetic variation at the DNA level in parental populations may increase the accuracy of predicting heterosis in crossbreeding.

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